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<p>(54) Title: DIAGNOSIS AND TREATMENT OF AUR-1 AND/OR AUR-2 RELATED DISORDERS (57) Abstract The present invention relates to AUR-1 and/or AUR-2 polypeptides, nucleic acids encoding such polypeptides, cells, tissues and animals containing such nucleic acids, antibodies to such polypeptides, assays utilizing such polypeptides, and methods relating to all of the foregoing. Methods for treatment, diagnosis, and screening are provided for AUR-1 and/or AUR-2 related diseases or conditions characterized by an abnormal interaction between an AUR-1 and/or AUR-2 polypeptide and an AUR-1 and/or AUR-2 binding partner.</p>		

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DESCRIPTIONDIAGNOSIS AND TREATMENT OF
AUR-1 and/or AUR-2 RELATED DISORDERSField of the Invention

The present invention relates to the novel protein termed AURORA ONE and AURORA TWO ("AUR-1 and AUR-2"), nucleotide sequences encoding AUR-1 and/or AUR-2, as well as various products and methods useful for the diagnosis and treatment of various AUR-1 and/or AUR-2 related diseases and conditions.

Background of the Invention

The following description of the background of the invention is provided to aid in understanding the invention but is not admitted to be prior art to the invention.

Cellular signal transduction is a fundamental mechanism whereby external stimuli that regulate diverse cellular processes are relayed to the interior of cells. One of the key biochemical mechanisms of signal transduction involves the reversible phosphorylation of proteins, which enables regulation of the activity of mature proteins by altering their structure and function.

The best characterized protein kinases in eukaryotes phosphorylate proteins on the alcohol moiety of serine, threonine and tyrosine residues. These

kinases largely fall into two groups, those specific for phosphorylating serines and threonines, and those specific for phosphorylating tyrosines. Some kinases, referred to as "dual specificity" kinases, are able to phosphorylate on tyrosine as well as serine/threonine residues.

Protein kinases can also be characterized by their location within the cell. Some kinases are transmembrane receptor-type proteins capable of directly altering their catalytic activity in response to the external environment such as the binding of a ligand. Others are non-receptor-type proteins lacking any transmembrane domain. They can be found in a variety of cellular compartments from the inner surface of the cell membrane to the nucleus.

Many kinases are involved in regulatory cascades wherein their substrates may include other kinases whose activities are regulated by their phosphorylation state. Ultimately the activity of some downstream effector is modulated by phosphorylation resulting from activation of such a pathway.

The serine/threonine kinase family includes members found at all steps of various signaling cascades, including those involved in controlling cell growth, migration, differentiation and secretion of hormones, phosphorylation of transcription factors resulting in altered gene expression, muscle contraction, glucose metabolism, control of cellular protein synthesis, and regulation of the cell cycle.

The present invention relates to AUR-1 and/or AUR-2 polypeptides, nucleic acids encoding such polypeptides, cells containing such nucleic acids, antibodies to such polypeptides, assays utilizing such polypeptides, and methods relating to all of the foregoing. The present invention is based upon the isolation and characterization of new proteins which we have designated AUR-1 and/or AUR-2. The polypeptides and nucleic acids may be produced using well known and standard synthesis techniques when given the sequences presented herein. AUR-1 and AUR-2 are related serine/threonine kinases with short N-terminal extensions. The Drosophila and yeast homologs appear to be involved in mitotic regulation. The human proteins appear to be involved in cancer and/or other signal transduction disorders. AUR1 RNA is broadly expressed in rapidly dividing cells, derived from both normal tumor tissues. AUR2 RNA however, is expressed in a more restricted pattern being low or absent in most normal tissues, and abundant in only a subset of tumor-derived cell lines, particularly those of colorectal origin. Both Auroral and Aurora2 show intermediate expression in fetal liver, adult testis, and thymus, suggestive of a normal role for these proteins in meiotic division. Both AUR1 and AUR2 appear to regulate nuclear division, with disruption of their signaling resulting in polyploid cells. This phenotype is likely due to chromosomal missegregation, as seen with their yeast homologue IPL1. Since polyploidy is a hallmark of tumor cells and in cells defective in the p53 tumor

suppressor, we are testing the role AUR1 and AUR2 in cellular transformation.

Primary sequence analysis of the human genes reveals that they contain a highly conserved C-terminal protein kinase domain and a weakly conserved N-terminal domain of 74 to 130 amino acids that may play a regulatory role or function as a substrate binding motif. The human genes also contain a cAMP/PKA phosphorylation site R/KR/KXS/T in the activation loop of the kinase domain, which suggests a regulatory pathway similar to the cell cycle regulated CDC2/CDK-related proteins.

Southern analysis with probes derived from the unique N-terminal regions of AUR1 and AUR2 indicate that they exist in as a single copy genes in human cells. However, under low stringency conditions, we were able to detect 1.3 kb and 3.2 kb SacI fragments which weakly hybridize to the AUR1 probe. Cloning and sequence analysis reveals this region to encode an intronless AUR1-related pseudogene (termed AUR3), with multiple frame shifts. Furthermore, immediately upstream of the AUR3 pseudogene is a region with complex inverted repeats predicted to form a very stable hairpin loop. AUR3 DNA sequence is homologous to AUR1 beginning from the first nucleotide of the AUR1 cDNA. Immediately upstream of this site, is the predicted hairpin loop of AUR3. We are currently characterizing AUR1 genomic clones to determine if the homology to AUR3 continues upstream from this nucleotide, and whether the AUR1 cDNA includes or be preceded by a similar hairpin loop.

The utility of the present invention includes the ability to screen for inhibitors of cell growth and to develop small molecule therapeutics for treating cancers.

Thus, in a first aspect the invention features
5 an isolated, enriched, or purified nucleic acid encoding a AUR-1 and/or AUR-2 polypeptide.

By "AUR-1 and/or AUR-2 polypeptide" is meant an amino acid sequence substantially similar to the sequence shown in SEQ ID NO:3 or SEQ ID NO:4, or
10 fragments thereof. A sequence that is substantially similar will preferably have at least 90% identity (more preferably at least 95% and most preferably 99-100%) to the sequence of SEQ ID NO:3 or SEQ ID NO:4.

By "identity" is meant a property of sequences
15 that measures their similarity or relationship. Identity is measured by dividing the number of identical residues by the total number of residues and multiplying the product by 100. Thus, two copies of exactly the same sequence have 100% identity, but sequences that are
20 less highly conserved and have deletions, additions, or replacements may have a lower degree of identity. Those skilled in the art will recognize that several computer programs are available for determining sequence identity.

By "isolated" in reference to nucleic acid is
25 meant a polymer of 6 (preferably 21, more preferably 39, most preferably 75) or more nucleotides conjugated to each other, including DNA or RNA that is isolated from a natural source or that is synthesized. In certain
30 embodiments of the invention longer nucleic acids are

preferred, for example those of 300, 600, 900 or more nucleotides and/or those having at least 50%, 60%, 75%, 90%, 95% or 99% identity to the full length sequence shown in SEQ ID NO:1 or SEQ ID NO:2. The isolated nucleic acid of the present invention is unique in the sense that it is not found in a pure or separated state in nature. Use of the term "isolated" indicates that a naturally occurring sequence has been removed from its normal cellular (i.e., chromosomal) environment. Thus, the sequence may be in a cell-free solution or placed in a different cellular environment. The term does not imply that the sequence is the only nucleotide chain present, but that it is essentially free (about 90 - 95% pure at least) of non-nucleotide material naturally associated with it and thus is meant to distinguished from isolated chromosomes.

By the use of the term "enriched" in reference to nucleic acid is meant that the specific DNA or RNA sequence constitutes a significantly higher fraction (2 - 5 fold) of the total DNA or RNA present in the cells or solution of interest than in normal or diseased cells or in the cells from which the sequence was taken. This could be caused by a person by preferential reduction in the amount of other DNA or RNA present, or by a preferential increase in the amount of the specific DNA or RNA sequence, or by a combination of the two. However, it should be noted that enriched does not imply that there are no other DNA or RNA sequences present, just that the relative amount of the sequence of interest has been significantly increased. The term significant here is used to indicate that the level of

increase is useful to the person making such an increase, and generally means an increase relative to other nucleic acids of about at least 2 fold, more preferably at least 5 to 10 fold or even more. The term also does not imply that there is no DNA or RNA from other sources. The other source DNA may, for example, comprise DNA from a yeast or bacterial genome, or a cloning vector such as pUC19. This term distinguishes from naturally occurring events, such as viral infection, or tumor type growths, in which the level of one mRNA may be naturally increased relative to other species of mRNA. That is, the term is meant to cover only those situations in which a person has intervened to elevate the proportion of the desired nucleic acid.

It is also advantageous for some purposes that a nucleotide sequence be in purified form. The term "purified" in reference to nucleic acid does not require absolute purity (such as a homogeneous preparation); instead, it represents an indication that the sequence is relatively purer than in the natural environment (compared to the natural level this level should be at least 2-5 fold greater, e.g., in terms of mg/ml). Individual clones isolated from a cDNA library may be purified to electrophoretic homogeneity. The claimed DNA molecules obtained from these clones could be obtained directly from total DNA or from total RNA. The cDNA clones are not naturally occurring, but rather are preferably obtained via manipulation of a partially purified naturally occurring substance (messenger RNA). The construction of a cDNA library from mRNA involves the creation of a synthetic substance (cDNA) and pure

individual cDNA clones can be isolated from the synthetic library by clonal selection of the cells carrying the cDNA library. Thus, the process which includes the construction of a cDNA library from mRNA and isolation of distinct cDNA clones yields an
5 approximately 10^6 -fold purification of the native message. Thus, purification of at least one order of magnitude, preferably two or three orders, and more preferably four or five orders of magnitude is expressly contemplated.

10 By "a AUR-1 and/or AUR-2 polypeptide" is meant 25 (preferably 30, more preferably 35, most preferably 40) or more contiguous amino acids set forth in the full length amino acid sequence of SEQ ID NO:3 or SEQ ID NO:4, or a functional derivative thereof as described
15 herein. In certain aspects, polypeptides of 100, 200, 300 or more amino acids are preferred. The AUR-1 and/or AUR-2 polypeptide can be encoded by a full-length nucleic acid sequence or any portion of the full-length nucleic acid sequence, so long as a functional activity
20 of the polypeptide is retained.

In preferred embodiments the isolated nucleic acid comprises, consists essentially of, or consists of a nucleic acid sequence set forth in the full length amino acid sequence of SEQ ID NO:3 or SEQ ID NO:4, a
25 functional derivative thereof, or encodes at least 25, 30, 35, 40, 50, 100, 200, or 300 contiguous amino acids thereof; the AUR-1 and/or AUR-2 polypeptide comprises, consists essentially of, or consists of at least 25, 30, 35, or 40 contiguous amino acids of a AUR-1 and/or AUR-2
30 polypeptide. The nucleic acid may be isolated from a

natural source by cDNA cloning or subtractive hybridization; the natural source may be mammalian (human) blood, semen, or tissue and the nucleic acid may be synthesized by the triester method or by using an automated DNA synthesizer. In yet other preferred
5 embodiments the nucleic acid is a conserved or unique region, for example those useful for the design of hybridization probes to facilitate identification and cloning of additional polypeptides, the design of PCR probes to facilitate cloning of additional polypeptides,
10 and obtaining antibodies to polypeptide regions. Examples of amino acid sequences of the present invention include the following amino acid sequences (the isolated, purified or enriched nucleic acids encoding them are also within the scope of the present
15 invention):

ENSYPWPYGRQ (SEQ ID NO:5)
CISGP (SEQ ID NO:6)
QFPO (SEQ ID NO:7)
VNSGQ (SEQ ID NO:8)
20 RKEPVTPSA-LV (SEQ ID NO:9)
LMSRSNVQPTAAP (SEQ ID NO:10)
VQNQKQKQLQATSVPH (SEQ ID NO:11)
PVSRLNNTQK (SEQ ID NO:12)
VMENSSGTPD (SEQ ID NO:13)
25 ILTRHFTID (SEQ ID NO:14)
SKQPLPSAPENNPEEQLASKQK (SEQ ID NO:15)

By "conserved nucleic acid regions", are meant regions present on two or more nucleic acids encoding a AUR-1 and/or AUR-2 polypeptide, to which a particular
30 nucleic acid sequence can hybridize under lower

stringency conditions. Examples of lower stringency conditions suitable for screening for nucleic acid encoding AUR-1 and/or AUR-2 polypeptides are provided in Abe, et al. J. Biol. Chem., 19:13361 (1992) (hereby incorporated by reference herein in its entirety, including any drawings). Preferably, conserved regions differ by no more than 5 out of 20 nucleotides.

By "unique nucleic acid region" is meant a sequence present in a full length nucleic acid coding for a AUR-1 and/or AUR-2 polypeptide that is not present in a sequence coding for any other naturally occurring polypeptide. Such regions preferably comprise 30 or 45 contiguous nucleotides present in the full length nucleic acid encoding a AUR-1 and/or AUR-2 polypeptide. In particular, a unique nucleic acid region is preferably of mammalian origin.

The invention also features a nucleic acid probe for the detection of a AUR-1 and/or AUR-2 polypeptide or nucleic acid encoding a AUR-1 and/or AUR-2 polypeptide in a sample. The nucleic acid probe contains nucleic acid that will hybridize to a sequence set forth in SEQ ID NO:1 or SEQ ID NO:2 or a functional derivative thereof.

In preferred embodiments the nucleic acid probe hybridizes to nucleic acid encoding at least 12, 75, 90, 105, 120, 150, 200, 250, 300 or 350 contiguous amino acids of the full-length sequence set forth in SEQ ID NO:1 or SEQ ID NO:2 or a functional derivative thereof. Various low or high stringency hybridization conditions may be used depending upon the specificity and selectivity desired. Under stringent hybridization

conditions only highly complementary nucleic acid sequences hybridize. Preferably, such conditions prevent hybridization of nucleic acids having 1 or 2 mismatches out of 20 contiguous nucleotides.

Methods for using the probes include detecting
5 the presence or amount of AUR-1 and/or AUR-2 RNA in a sample by contacting the sample with a nucleic acid probe under conditions such that hybridization occurs and detecting the presence or amount of the probe bound to AUR-1 and/or AUR-2 RNA. The nucleic acid duplex
10 formed between the probe and a nucleic acid sequence coding for a AUR-1 and/or AUR-2 polypeptide may be used in the identification of the sequence of the nucleic acid detected (for example see, Nelson et al., in Nonisotopic DNA Probe Techniques, p. 275 Academic Press,
15 San Diego (Kricka, ed., 1992) hereby incorporated by reference herein in its entirety, including any drawings). Kits for performing such methods may be constructed to include a container means having disposed therein a nucleic acid probe.

20 The invention also features recombinant nucleic acid, preferably in a cell or an organism. The recombinant nucleic acid may contain a sequence set forth in SEQ ID NO:1 or SEQ ID NO:2 or a functional derivative thereof and a vector or a promoter effective
25 to initiate transcription in a host cell. The recombinant nucleic acid can alternatively contain a transcriptional initiation region functional in a cell, a sequence complimentary to an RNA sequence encoding a AUR-1 and/or AUR-2 polypeptide and a transcriptional
30 termination region functional in a cell.

In another aspect the invention features an isolated, enriched, or purified AUR-1 and/or AUR-2 polypeptide.

By "isolated" in reference to a polypeptide is meant a polymer of 2 (preferably 7, more preferably 13, most preferably 25) or more amino acids conjugated to each other, including polypeptides that are isolated from a natural source or that are synthesized. In certain aspects longer polypeptides are preferred, such as those with 402, 407, 413, or 425 contiguous amino acids set forth in SEQ ID NO:3 or SEQ ID NO:4. The isolated polypeptides of the present invention are unique in the sense that they are not found in a pure or separated state in nature. Use of the term "isolated" indicates that a naturally occurring sequence has been removed from its normal cellular environment. Thus, the sequence may be in a cell-free solution or placed in a different cellular environment. The term does not imply that the sequence is the only amino acid chain present, but that it is essentially free (about 90 - 95% pure at least) of non-amino acid material naturally associated with it.

By the use of the term "enriched" in reference to a polypeptide is meant that the specific amino acid sequence constitutes a significantly higher fraction (2 - 5 fold) of the total of amino acids present in the cells or solution of interest than in normal or diseased cells or in the cells from which the sequence was taken. This could be caused by a person by preferential reduction in the amount of other amino acids present, or by a preferential increase in the amount of the specific

amino acid sequence of interest, or by a combination of the two. However, it should be noted that enriched does not imply that there are no other amino acid sequences present, just that the relative amount of the sequence of interest has been significantly increased. The term
5 significant here is used to indicate that the level of increase is useful to the person making such an increase, and generally means an increase relative to other amino acids of about at least 2 fold, more preferably at least 5 to 10 fold or even more. The term
10 also does not imply that there is no amino acid from other sources. The other source amino acid may, for example, comprise amino acid encoded by a yeast or bacterial genome, or a cloning vector such as pUC19. The term is meant to cover only those situations in
15 which man has intervened to elevate the proportion of the desired nucleic acid.

It is also advantageous for some purposes that an amino acid sequence be in purified form. The term "purified" in reference to a polypeptide does not
20 require absolute purity (such as a homogeneous preparation); instead, it represents an indication that the sequence is relatively purer than in the natural environment (compared to the natural level this level should be at least 2-5 fold greater, e.g., in terms of
25 mg/ml). Purification of at least one order of magnitude, preferably two or three orders, and more preferably four or five orders of magnitude is expressly contemplated. The substance is preferably free of contamination at a functionally significant level, for
30 example 90%, 95%, or 99% pure.

In preferred embodiments the AUR-1 and/or AUR-2 polypeptide contains at least 25, 30, 35, 40, 50, 100, 150, 200, 250, 300, or 350 contiguous amino acids of the full-length sequence set forth in SEQ ID NO:3 or SEQ ID NO:4, or a functional derivative thereof.

5 In yet another aspect the invention features an antibody (e.g., a monoclonal or polyclonal antibody) having specific binding affinity to a AUR-1 and/or AUR-2 polypeptide. The antibody contains a sequence of amino acids that is able to specifically bind to a AUR-1
10 and/or AUR-2 polypeptide. By "specific binding affinity" is meant that the antibody binds to AUR-1 and/or AUR-2 polypeptides with greater affinity than it binds to other polypeptides under specified conditions.

Antibodies having specific binding affinity to
15 a AUR-1 and/or AUR-2 polypeptide may be used in methods for detecting the presence and/or amount of a AUR-1 and/or AUR-2 polypeptide in a sample by contacting the sample with the antibody under conditions such that an immunocomplex forms and detecting the presence and/or
20 amount of the antibody conjugated to the AUR-1 and/or AUR-2 polypeptide. Diagnostic kits for performing such methods may be constructed to include a first container means containing the antibody and a second container means having a conjugate of a binding partner of the
25 antibody and a label.

In another aspect the invention features a hybridoma which produces an antibody having specific binding affinity to a AUR-1 and/or AUR-2 polypeptide. By "hybridoma" is meant an immortalized cell line which
30 is capable of secreting an antibody, for example a AUR-1

and/or AUR-2 antibody. In preferred embodiments the AUR-1 and/or AUR-2 antibody comprises a sequence of amino acids that is able to specifically bind a AUR-1 and/or AUR-2 polypeptide.

In another aspect, the invention describes a polypeptide comprising a recombinant AUR-1 and/or AUR-2 polypeptide or a unique fragment thereof. By "unique fragment," is meant an amino acid sequence present in a full-length AUR-1 and/or AUR-2 polypeptide that is not present in any other naturally occurring polypeptide. Preferably, such a sequence comprises 6 contiguous amino acids present in the full sequence. More preferably, such a sequence comprises 12 contiguous amino acids present in the full sequence. Even more preferably, such a sequence comprises 18 contiguous amino acids present in the full sequence.

By "recombinant AUR-1 and/or AUR-2 polypeptide" is meant to include a polypeptide produced by recombinant DNA techniques such that it is distinct from a naturally occurring polypeptide either in its location (e.g., present in a different cell or tissue than found in nature), purity or structure. Generally, such a recombinant polypeptide will be present in a cell in an amount different from that normally observed in nature.

In another aspect, the invention describes a recombinant cell or tissue containing a purified nucleic acid coding for a AUR-1 and/or AUR-2 polypeptide. In such cells, the nucleic acid may be under the control of its genomic regulatory elements, or may be under the control of exogenous regulatory elements including an

exogenous promoter. By "exogenous" it is meant a promoter that is not normally coupled *in vivo* transcriptionally to the coding sequence for the AUR-1 and/or AUR-2 polypeptide.

In another aspect, the invention features a
5 AUR-1 and/or AUR-2 polypeptide binding agent able to bind to a AUR-1 and/or AUR-2 polypeptide. The binding agent is preferably a purified antibody which recognizes an epitope present on a AUR-1 and/or AUR-2 polypeptide. Other binding agents include molecules which bind to the
10 AUR-1 and/or AUR-2 polypeptide and analogous molecules which bind to a AUR-1 and/or AUR-2 polypeptide. Such binding agents may be identified by using assays that measure AUR-1 and/or AUR-2 binding partner activity, such as those that measure PDGFR activity.

15 By "purified" in reference to an antibody is meant that the antibody is distinct from naturally occurring antibody, such as in a purified form. Preferably, the antibody is provided as a homogeneous preparation by standard techniques. Uses of antibodies
20 to the cloned polypeptide include those to be used as therapeutics, or as diagnostic tools.

The invention features a method for screening for human cells containing a AUR-1 and/or AUR-2 polypeptide or an equivalent sequence. The method
25 involves identifying the novel polypeptide in human cells using techniques that are routine and standard in the art, such as those described herein for identifying AUR-1 and/or AUR-2 (e.g., cloning, Southern or Northern blot analysis, in situ hybridization, PCR amplification,
30 etc.).

The invention also features methods of screening human cells for binding partners of AUR-1 and/or AUR-2 polypeptides and screening other organisms for AUR-1 and/or AUR-2 or the corresponding binding partner. The present invention also features the
5 purified, isolated or enriched versions of the peptides identified by the methods described above.

In another aspect, the invention provides an assay to identify agents capable of interfering with the interaction between AUR-1 and/or AUR-2 and a AUR-1
10 and/or AUR-2 binding partner. Such assays may be performed in vitro or in vivo and are described in detail herein or can be obtained by modifying existing assays, such as the growth assay described in Serial No. 08/487,088, filed June 7, 1995, (incorporated herein by
15 reference including any drawings) or the assays described in Serial No. 60/005,167, filed October 13, 1995 (incorporated herein by reference including any drawings). Another assay which could be modified to use the genes of the present invention are described in
20 International Application No. WO 94/23039, published October 13, 1994. Other possibilities include detecting kinase activity in an autophosphorylation assay or testing for kinase activity on standard substrates such as histones, myelin basic protein, gamma tubulin, or
25 centrosomal proteins. Binding partners may be identified by putting the N-terminal portion of the protein into a two-hybrid screen or detecting phosphotyrosine of a dual specificity kinase. Fields and Song, U.S. Patent No. 5,283,173, issued February 1,
30 1994 and is incorporated by reference herein.

One means by which inhibitors of Aurora activity may be defined is a screening system using a temperature-sensitive yeast mutant as described by Chan and Botstein (Genetics 135:677-691, 1993); see also Francisco et al., Mol. Cell. Bio. 14(7):4731-4740, 1994) both of which are hereby incorporated herein by reference in their entirety including any drawings. Briefly, yeast strain CCY72-3D-1 (ipl 1-2), which expresses a temperature sensitive form of the yeast homologue of Aurora (ipl1) while viable at 26°C is incapable of growth at 37°C. Transfection of this strain with an expression plasmid containing a hybrid Aurora gene consisting of the N-terminal portion of ipl1, containing the putative substrate interaction domain(s)), and the C-terminal portion of Auroras 1 or 2, containing the catalytic domain, overcomes this sensitivity to growth temperature. The Aurora-expressing yeast strain is then grown at 37°C in the presence of a test substance. No growth will be evident in the presence of substances that inhibit Aurora catalytic function. Potential inhibitors include small molecular weight chemicals and/or natural products isolation from diverse organisms such as fungi, marine organisms, plants, etc.

The summary of the invention described above is non-limiting and other features and advantages of the invention will be apparent from the following description of the preferred embodiments, and from the claims.

Description of the Preferred Embodiments

The present invention relates to AUR-1 and/or AUR-2 polypeptides, nucleic acids encoding such polypeptides, cells, tissues and animals containing such nucleic acids, antibodies to such polypeptides, assays utilizing such polypeptides, and methods relating to all of the foregoing.

I. Nucleic Acid Encoding AUR-1 and/or AUR-2 Polypeptides.

Included within the scope of this invention are the functional equivalents of the herein-described isolated nucleic acid molecules. The degeneracy of the genetic code permits substitution of certain codons by other codons which specify the same amino acid and hence would give rise to the same protein. The nucleic acid sequence can vary substantially since, with the exception of methionine and tryptophan, the known amino acids can be coded for by more than one codon. Thus, portions or all of the AUR-1 and/or AUR-2 gene could be synthesized to give a nucleic acid sequence significantly different from that shown in SEQ ID NO:1 or SEQ ID NO:2. The encoded amino acid sequence thereof would, however, be preserved.

In addition, the nucleic acid sequence may comprise a nucleotide sequence which results from the addition, deletion or substitution of at least one nucleotide to the 5'-end and/or the 3'-end of the nucleic acid formula shown in SEQ ID NO:1 or SEQ ID NO:2 or a derivative thereof. Any nucleotide or polynucleotide may be used in this regard, provided that its addition, deletion or substitution does not alter

the amino acid sequence of SEQ ID NO:3 or SEQ ID NO:4 which is encoded by the nucleotide sequence. For example, the present invention is intended to include any nucleic acid sequence resulting from the addition of ATG as an initiation codon at the 5'-end of the
5 inventive nucleic acid sequence or its derivative, or from the addition of TTA, TAG or TGA as a termination codon at the 3'-end of the inventive nucleotide sequence or its derivative. Moreover, the nucleic acid molecule of the present invention may, as necessary, have
10 restriction endonuclease recognition sites added to its 5'-end and/or 3'-end.

Such functional alterations of a given nucleic acid sequence afford an opportunity to promote secretion and/or processing of heterologous proteins encoded by
15 foreign nucleic acid sequences fused thereto. All variations of the nucleotide sequence of the AUR-1 and/or AUR-2 genes and fragments thereof permitted by the genetic code are, therefore, included in this invention.

20 Further, it is possible to delete codons or to substitute one or more codons by codons other than degenerate codons to produce a structurally modified polypeptide, but one which has substantially the same utility or activity of the polypeptide produced by the
25 unmodified nucleic acid molecule. As recognized in the art, the two polypeptides are functionally equivalent, as are the two nucleic acid molecules which give rise to their production, even though the differences between the nucleic acid molecules are not related to degeneracy
30 of the genetic code.

II. A Nucleic Acid Probe for the Detection of AUR-1 and/or AUR-2.

A nucleic acid probe of the present invention may be used to probe an appropriate chromosomal or cDNA library by usual hybridization methods to obtain another
5 nucleic acid molecule of the present invention. A chromosomal DNA or cDNA library may be prepared from appropriate cells according to recognized methods in the art (cf. "Molecular Cloning: A Laboratory Manual", second edition, edited by Sambrook, Fritsch, & Maniatis,
10 Cold Spring Harbor Laboratory, 1989).

In the alternative, chemical synthesis is carried out in order to obtain nucleic acid probes having nucleotide sequences which correspond to N-terminal and C-terminal portions of the amino acid
15 sequence of the polypeptide of interest. Thus, the synthesized nucleic acid probes may be used as primers in a polymerase chain reaction (PCR) carried out in accordance with recognized PCR techniques, essentially according to PCR Protocols, "A Guide to Methods and
20 Applications", edited by Michael et al., Academic Press, 1990, utilizing the appropriate chromosomal or cDNA library to obtain the fragment of the present invention.

One skilled in the art can readily design such probes based on the sequence disclosed herein using
25 methods of computer alignment and sequence analysis known in the art (cf. "Molecular Cloning: A Laboratory Manual", second edition, edited by Sambrook, Fritsch, & Maniatis, Cold Spring Harbor Laboratory, 1989). The hybridization probes of the present invention can be
30 labeled by standard labeling techniques such as with a

radiolabel, enzyme label, fluorescent label, biotin-
avidin label, chemiluminescence, and the like. After
hybridization, the probes may be visualized using known
methods.

The nucleic acid probes of the present
5 invention include RNA, as well as DNA probes, such
probes being generated using techniques known in the
art. The nucleic acid probe may be immobilized on a
solid support. Examples of such solid supports include,
but are not limited to, plastics such as polycarbonate,
10 complex carbohydrates such as agarose and sepharose, and
acrylic resins, such as polyacrylamide and latex beads.
Techniques for coupling nucleic acid probes to such
solid supports are well known in the art.

The test samples suitable for nucleic acid
15 probing methods of the present invention include, for
example, cells or nucleic acid extracts of cells, or
biological fluids. The sample used in the above-
described methods will vary based on the assay format,
the detection method and the nature of the tissues,
20 cells or extracts to be assayed. Methods for preparing
nucleic acid extracts of cells are well known in the art
and can be readily adapted in order to obtain a sample
which is compatible with the method utilized.

25 III. A Probe Based Method And Kit For Detecting AUR-1 and/or AUR-2.

One method of detecting the presence of AUR-1
and/or AUR-2 in a sample comprises (a) contacting said
sample with the above-described nucleic acid probe,
30 under conditions such that hybridization occurs, and (b)

detecting the presence of said probe bound to said nucleic acid molecule. One skilled in the art would select the nucleic acid probe according to techniques known in the art as described above. Samples to be tested include but should not be limited to RNA samples of human tissue.

A kit for detecting the presence of AUR-1 and/or AUR-2 in a sample comprises at least one container means having disposed therein the above-described nucleic acid probe. The kit may further comprise other containers comprising one or more of the following: wash reagents and reagents capable of detecting the presence of bound nucleic acid probe. Examples of detection reagents include, but are not limited to radiolabelled probes, enzymatic labeled probes (horseradish peroxidase, alkaline phosphatase), and affinity labeled probes (biotin, avidin, or streptavidin).

In detail, a compartmentalized kit includes any kit in which reagents are contained in separate containers. Such containers include small glass containers, plastic containers or strips of plastic or paper. Such containers allow the efficient transfer of reagents from one compartment to another compartment such that the samples and reagents are not cross-contaminated and the agents or solutions of each container can be added in a quantitative fashion from one compartment to another. Such containers will include a container which will accept the test sample, a container which contains the probe or primers used in the assay, containers which contain wash reagents (such

as phosphate buffered saline, Tris-buffers, and the like), and containers which contain the reagents used to detect the hybridized probe, bound antibody, amplified product, or the like. One skilled in the art will readily recognize that the nucleic acid probes described
5 in the present invention can readily be incorporated into one of the established kit formats which are well known in the art.

**IV. DNA Constructs Comprising a AUR-1 and/or AUR-2
10 Nucleic Acid Molecule and Cells Containing These
Constructs.**

The present invention also relates to a recombinant DNA molecule comprising, 5' to 3', a promoter effective to initiate transcription in a host
15 cell and the above-described nucleic acid molecules. In addition, the present invention relates to a recombinant DNA molecule comprising a vector and an above-described nucleic acid molecules. The present invention also relates to a nucleic acid molecule comprising a
20 transcriptional region functional in a cell, a sequence complimentary to an RNA sequence encoding an amino acid sequence corresponding to the above-described polypeptide, and a transcriptional termination region functional in said cell. The above-described molecules
25 may be isolated and/or purified DNA molecules..

The present invention also relates to a cell or organism that contains an above-described nucleic acid molecule and thereby is capable of expressing a peptide. The polypeptide may be purified from cells
30 which have been altered to express the polypeptide. A

cell is said to be "altered to express a desired polypeptide" when the cell, through genetic manipulation, is made to produce a protein which it normally does not produce or which the cell normally produces at lower levels. One skilled in the art can readily adapt procedures for introducing and expressing either genomic, cDNA, or synthetic sequences into either eukaryotic or prokaryotic cells.

A nucleic acid molecule, such as DNA, is said to be "capable of expressing" a polypeptide if it contains nucleotide sequences which contain transcriptional and translational regulatory information and such sequences are "operably linked" to nucleotide sequences which encode the polypeptide. An operable linkage is a linkage in which the regulatory DNA sequences and the DNA sequence sought to be expressed are connected in such a way as to permit gene sequence expression. The precise nature of the regulatory regions needed for gene sequence expression may vary from organism to organism, but shall in general include a promoter region which, in prokaryotes, contains both the promoter (which directs the initiation of RNA transcription) as well as the DNA sequences which, when transcribed into RNA, will signal synthesis initiation. Such regions will normally include those 5'-non-coding sequences involved with initiation of transcription and translation, such as the TATA box, capping sequence, CAAT sequence, and the like.

If desired, the non-coding region 3' to the sequence encoding an AUR-1 and/or AUR-2 gene may be obtained by the above-described methods. This region

may be retained for its transcriptional termination regulatory sequences, such as termination and polyadenylation. Thus, by retaining the 3'-region naturally contiguous to the DNA sequence encoding an AUR-1 and/or AUR-2 gene, the transcriptional termination
5 signals may be provided. Where the transcriptional termination signals are not satisfactorily functional in the expression host cell, then a 3' region functional in the host cell may be substituted.

Two DNA sequences (such as a promoter region
10 sequence and an AUR-1 and/or AUR-2 sequence) are said to be operably linked if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region sequence to
15 direct the transcription of an AUR-1 and/or AUR-2 gene sequence, or (3) interfere with the ability of the an AUR-1 and/or AUR-2 gene sequence to be transcribed by the promoter region sequence. Thus, a promoter region would be operably linked to a DNA sequence if the
20 promoter were capable of effecting transcription of that DNA sequence. Thus, to express an AUR-1 and/or AUR-2 gene, transcriptional and translational signals recognized by an appropriate host are necessary.

The present invention encompasses the
25 expression of the AUR-1 and/or AUR-2 gene (or a functional derivative thereof) in either prokaryotic or eukaryotic cells. Prokaryotic hosts are, generally, very efficient and convenient for the production of recombinant proteins and are, therefore, one type of
30 preferred expression system for the AUR-1 and/or AUR-2

gene. Prokaryotes most frequently are represented by various strains of *E. coli*. However, other microbial strains may also be used, including other bacterial strains.

In prokaryotic systems, plasmid vectors that contain replication sites and control sequences derived from a species compatible with the host may be used. Examples of suitable plasmid vectors may include pBR322, pUC118, pUC119 and the like; suitable phage or bacteriophage vectors may include γ gt10, γ gt11 and the like; and suitable virus vectors may include pMAM-neo, pKRC and the like. Preferably, the selected vector of the present invention has the capacity to replicate in the selected host cell.

Recognized prokaryotic hosts include bacteria such as *E. coli*, *Bacillus*, *Streptomyces*, *Pseudomonas*, *Salmonella*, *Serratia*, and the like. However, under such conditions, the peptide will not be glycosylated. The prokaryotic host must be compatible with the replicon and control sequences in the expression plasmid.

To express AUR-1 and/or AUR-2 (or a functional derivative thereof) in a prokaryotic cell, it is necessary to operably link the AUR-1 and/or AUR-2 sequence to a functional prokaryotic promoter. Such promoters may be either constitutive or, more preferably, regulatable (i.e., inducible or derepressible). Examples of constitutive promoters include the *int* promoter of bacteriophage λ , the *bla* promoter of the β -lactamase gene sequence of pBR322, and the CAT promoter of the chloramphenicol acetyl transferase gene sequence of pPR325, and the like.

Examples of inducible prokaryotic promoters include the major right and left promoters of bacteriophage λ (P_L and P_R), the trp, recA, lacZ, lacI, and gal promoters of E. coli, the α -amylase (Ulmanen et al., J. Bacteriol. 162:176-182(1985)) and the ζ -28-specific promoters of B. subtilis (Gilman et al., Gene sequence 32:11-20(1984)),
5 the promoters of the bacteriophages of Bacillus (Gryczan, In: The Molecular Biology of the Bacilli, Academic Press, Inc., NY (1982)), and Streptomyces promoters (Ward et al., Mol. Gen. Genet. 203:468-
10 478(1986)). Prokaryotic promoters are reviewed by Glick (J. Ind. Microbiol. 1:277-282(1987)); Cenatiempo (Biochimie 68:505-516(1986)); and Gottesman (Ann. Rev. Genet. 18:415-442 (1984)).

Proper expression in a prokaryotic cell also
15 requires the presence of a ribosome binding site upstream of the gene sequence-encoding sequence. Such ribosome binding sites are disclosed, for example, by Gold et al. (Ann. Rev. Microbiol. 35:365-404(1981)). The selection of control sequences, expression vectors,
20 transformation methods, and the like, are dependent on the type of host cell used to express the gene. As used herein, "cell", "cell line", and "cell culture" may be used interchangeably and all such designations include progeny. Thus, the words "transformants" or
25 "transformed cells" include the primary subject cell and cultures derived therefrom, without regard to the number of transfers. It is also understood that all progeny may not be precisely identical in DNA content, due to deliberate or inadvertent mutations. However, as

defined, mutant progeny have the same functionality as that of the originally transformed cell.

Host cells which may be used in the expression systems of the present invention are not strictly limited, provided that they are suitable for use in the expression of the AUR-1 and/or AUR-2 peptide of interest. Suitable hosts may often include eukaryotic cells. Preferred eukaryotic hosts include, for example, yeast, fungi, insect cells, mammalian cells either in vivo, or in tissue culture. Mammalian cells which may be useful as hosts include HeLa cells, cells of fibroblast origin such as VERO or CHO-K1, or cells of lymphoid origin and their derivatives. Preferred mammalian host cells include SP2/0 and J558L, as well as neuroblastoma cell lines such as IMR 332 which may provide better capacities for correct post-translational processing.

In addition, plant cells are also available as hosts, and control sequences compatible with plant cells are available, such as the cauliflower mosaic virus 35S and 19S, and nopaline synthase promoter and polyadenylation signal sequences. Another preferred host is an insect cell, for example the *Drosophila* larvae. Using insect cells as hosts, the *Drosophila* alcohol dehydrogenase promoter can be used. Rubin, Science 240:1453-1459(1988). Alternatively, baculovirus vectors can be engineered to express large amounts of AUR-1 and/or AUR-2 in insects cells (Jasny, Science 238:1653 (1987); Miller et al., In: Genetic Engineering (1986), Setlow, J.K., et al., eds., Plenum, Vol. 8, pp. 277-297).

Any of a series of yeast gene sequence expression systems can be utilized which incorporate promoter and termination elements from the actively expressed gene sequences coding for glycolytic enzymes are produced in large quantities when yeast are grown in mediums rich in glucose. Known glycolytic gene sequences can also provide very efficient transcriptional control signals. Yeast provides substantial advantages in that it can also carry out post-translational peptide modifications. A number of recombinant DNA strategies exist which utilize strong promoter sequences and high copy number of plasmids which can be utilized for production of the desired proteins in yeast. Yeast recognizes leader sequences on cloned mammalian gene sequence products and secretes peptides bearing leader sequences (i.e., pre-peptides). For a mammalian host, several possible vector systems are available for the expression of AUR-1 and/or AUR-2.

A wide variety of transcriptional and translational regulatory sequences may be employed, depending upon the nature of the host. The transcriptional and translational regulatory signals may be derived from viral sources, such as adenovirus, bovine papilloma virus, cytomegalovirus, simian virus, or the like, where the regulatory signals are associated with a particular gene sequence which has a high level of expression. Alternatively, promoters from mammalian expression products, such as actin, collagen, myosin, and the like, may be employed. Transcriptional initiation regulatory signals may be selected which allow for repression or activation, so that expression

of the gene sequences can be modulated. Of interest are regulatory signals which are temperature-sensitive so that by varying the temperature, expression can be repressed or initiated, or are subject to chemical (such as metabolite) regulation.

5 Expression of AUR-1 and/or AUR-2 in eukaryotic hosts requires the use of eukaryotic regulatory regions. Such regions will, in general, include a promoter region sufficient to direct the initiation of RNA synthesis. Preferred eukaryotic promoters include, for example, the
10 promoter of the mouse metallothionein I gene sequence (Hamer et al., J. Mol. Appl. Gen. 1:273-288(1982)); the TK promoter of Herpes virus (McKnight, Cell 31:355-365 (1982)); the SV40 early promoter (Benoist et al., Nature (London) 290:304-310(1981)); the yeast gal4 gene
15 sequence promoter (Johnston et al., Proc. Natl. Acad. Sci. (USA) 79:6971-6975(1982); Silver et al., Proc. Natl. Acad. Sci. (USA) 81:5951-5955 (1984)).

 Translation of eukaryotic mRNA is initiated at the codon which encodes the first methionine. For this
20 reason, it is preferable to ensure that the linkage between a eukaryotic promoter and a DNA sequence which encodes AUR-1 and/or AUR-2 (or a functional derivative thereof) does not contain any intervening codons which are capable of encoding a methionine (i.e., AUG). The
25 presence of such codons results either in a formation of a fusion protein (if the AUG codon is in the same reading frame as the AUR-1 and/or AUR-2 coding sequence) or a frame-shift mutation (if the AUG codon is not in the same reading frame as the AUR-1 and/or AUR-2 coding
30 sequence).

A AUR-1 and/or AUR-2 nucleic acid molecule and an operably linked promoter may be introduced into a recipient prokaryotic or eukaryotic cell either as a nonreplicating DNA (or RNA) molecule, which may either be a linear molecule or, more preferably, a closed covalent circular molecule. Since such molecules are incapable of autonomous replication, the expression of the gene may occur through the transient expression of the introduced sequence. Alternatively, permanent expression may occur through the integration of the introduced DNA sequence into the host chromosome.

A vector may be employed which is capable of integrating the desired gene sequences into the host cell chromosome. Cells which have stably integrated the introduced DNA into their chromosomes can be selected by also introducing one or more markers which allow for selection of host cells which contain the expression vector. The marker may provide for prototrophy to an auxotrophic host, biocide resistance, e.g., antibiotics, or heavy metals, such as copper, or the like. The selectable marker gene sequence can either be directly linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transfection. Additional elements may also be needed for optimal synthesis of single chain binding protein mRNA. These elements may include splice signals, as well as transcription promoters, enhancers, and termination signals. cDNA expression vectors incorporating such elements include those described by Okayama, Molec. Cell. Biol. 3:280 (1983).

The introduced nucleic acid molecule can be incorporated into a plasmid or viral vector capable of autonomous replication in the recipient host. Any of a wide variety of vectors may be employed for this purpose. Factors of importance in selecting a particular plasmid or viral vector include: the ease with which recipient cells that contain the vector may be recognized and selected from those recipient cells which do not contain the vector; the number of copies of the vector which are desired in a particular host; and whether it is desirable to be able to "shuttle" the vector between host cells of different species.

Preferred prokaryotic vectors include plasmids such as those capable of replication in *E. coli* (such as, for example, pBR322, ColE1, pSC101, pACYC 184, nVX). Such plasmids are, for example, disclosed by Sambrook (cf. "Molecular Cloning: A Laboratory Manual", second edition, edited by Sambrook, Fritsch, & Maniatis, Cold Spring Harbor Laboratory, (1989)). *Bacillus* plasmids include pC194, pC221, pT127, and the like. Such plasmids are disclosed by Gryczan (In: The Molecular Biology of the Bacilli, Academic Press, NY (1982), pp. 307-329). Suitable *Streptomyces* plasmids include p1J101 (Kendall et al., J. Bacteriol. 169:4177-4183 (1987)), and streptomyces bacteriophages such as ϕ C31 (Chater et al., In: Sixth International Symposium on Actinomycetales Biology, Akademiai Kiado, Budapest, Hungary (1986), pp. 45-54). *Pseudomonas* plasmids are reviewed by John et al. (Rev. Infect. Dis. 8:693-704 (1986)), and Izaki (Jpn. J. Bacteriol. 33:729-742 (1978)).

Preferred eukaryotic plasmids include, for example, BPV, vaccinia, SV40, 2-micron circle, and the like, or their derivatives. Such plasmids are well known in the art (Botstein et al., Miami Wntr. Symp. 19:265-274(1982); Broach, In: "The Molecular Biology of the Yeast *Saccharomyces*: Life Cycle and Inheritance", Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, p. 445-470 (1981); Broach, Cell 28:203-204 (1982); Bollon et al., J. Clin. Hematol. Oncol. 10:39-48 (1980); Maniatis, In: Cell Biology: A Comprehensive Treatise, Vol. 3, Gene Sequence Expression, Academic Press, NY, pp. 563-608(1980).

Once the vector or nucleic acid molecule containing the construct(s) has been prepared for expression, the DNA construct(s) may be introduced into an appropriate host cell by any of a variety of suitable means, i.e., transformation, transfection, conjugation, protoplast fusion, electroporation, particle gun technology, calcium phosphate-precipitation, direct microinjection, and the like. After the introduction of the vector, recipient cells are grown in a selective medium, which selects for the growth of vector-containing cells. Expression of the cloned gene molecule(s) results in the production of AUR-1 and/or AUR-2 or fragments thereof. This can take place in the transformed cells as such, or following the induction of these cells to differentiate (for example, by administration of bromodeoxyuracil to neuroblastoma cells or the like). A variety of incubation conditions can be used to form the peptide of the present

invention. The most preferred conditions are those which mimic physiological conditions.

V. Purified AUR-1 and/or AUR-2 Polypeptides

A variety of methodologies known in the art can be utilized to obtain the peptide of the present invention. The peptide may be purified from tissues or cells which naturally produce the peptide. Alternatively, the above-described isolated nucleic acid fragments could be used to expressed the AUR-1 and/or AUR-2 protein in any organism. The samples of the present invention include cells, protein extracts or membrane extracts of cells, or biological fluids. The sample will vary based on the assay format, the detection method and the nature of the tissues, cells or extracts used as the sample.

Any eukaryotic organism can be used as a source for the peptide of the invention, as long as the source organism naturally contains such a peptide. As used herein, "source organism" refers to the original organism from which the amino acid sequence of the subunit is derived, regardless of the organism the subunit is expressed in and ultimately isolated from.

One skilled in the art can readily follow known methods for isolating proteins in order to obtain the peptide free of natural contaminants. These include, but are not limited to: size-exclusion chromatography, HPLC, ion-exchange chromatography, and immuno-affinity chromatography.

VI. An Antibody Having Binding Affinity To A AUR-1 and/or AUR-2 Polypeptide And A Hybridoma Containing the Antibody.

The present invention relates to an antibody having binding affinity to a AUR-1 and/or AUR-2 polypeptide. The polypeptide may have the amino acid sequence set forth in SEQ ID NO:3 or SEQ ID NO:4, or functional derivative thereof, or at least 9 contiguous amino acids thereof (preferably, at least 20, 30, 35, or 40 contiguous amino acids thereof).

The present invention also relates to an antibody having specific binding affinity to an AUR-1 and/or AUR-2 polypeptide. Such an antibody may be isolated by comparing its binding affinity to a AUR-1 and/or AUR-2 polypeptide with its binding affinity to another polypeptide. Those which bind selectively to AUR-1 and/or AUR-2 would be chosen for use in methods requiring a distinction between AUR-1 and/or AUR-2 and other polypeptides. Such methods could include, but should not be limited to, the analysis of altered AUR-1 and/or AUR-2 expression in tissue containing other polypeptides.

The AUR-1 and/or AUR-2 proteins of the present invention can be used in a variety of procedures and methods, such as for the generation of antibodies, for use in identifying pharmaceutical compositions, and for studying DNA/protein interaction.

The AUR-1 and/or AUR-2 peptide of the present invention can be used to produce antibodies or hybridomas. One skilled in the art will recognize that if an antibody is desired, such a peptide would be

generated as described herein and used as an immunogen. The antibodies of the present invention include monoclonal and polyclonal antibodies, as well fragments of these antibodies, and humanized forms. Humanized forms of the antibodies of the present invention may be generated using one of the procedures known in the art such as chimerization or CDR grafting. The present invention also relates to a hybridoma which produces the above-described monoclonal antibody, or binding fragment thereof. A hybridoma is an immortalized cell line which is capable of secreting a specific monoclonal antibody.

In general, techniques for preparing monoclonal antibodies and hybridomas are well known in the art (Campbell, "Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology," Elsevier Science Publishers, Amsterdam, The Netherlands (1984); St. Groth et al., J. Immunol. Methods 35:1-21(1980)). Any animal (mouse, rabbit, and the like) which is known to produce antibodies can be immunized with the selected polypeptide. Methods for immunization are well known in the art. Such methods include subcutaneous or intraperitoneal injection of the polypeptide. One skilled in the art will recognize that the amount of polypeptide used for immunization will vary based on the animal which is immunized, the antigenicity of the polypeptide and the site of injection.

The polypeptide may be modified or administered in an adjuvant in order to increase the peptide antigenicity. Methods of increasing the antigenicity of a polypeptide are well known in the art.

Such procedures include coupling the antigen with a heterologous protein (such as globulin or β -galactosidase) or through the inclusion of an adjuvant during immunization.

For monoclonal antibodies, spleen cells from
5 the immunized animals are removed, fused with myeloma cells, such as SP2/0-Ag14 myeloma cells, and allowed to become monoclonal antibody producing hybridoma cells. Any one of a number of methods well known in the art can be used to identify the hybridoma cell which produces an
10 antibody with the desired characteristics. These include screening the hybridomas with an ELISA assay, western blot analysis, or radioimmunoassay (Lutz et al., Exp. Cell Res. 175:109-124(1988)). Hybridomas secreting the desired antibodies are cloned and the class and subclass
15 is determined using procedures known in the art (Campbell, "Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology", supra (1984)).

For polyclonal antibodies, antibody containing
20 antisera is isolated from the immunized animal and is screened for the presence of antibodies with the desired specificity using one of the above-described procedures. The above-described antibodies may be detectably labeled. Antibodies can be detectably labeled through
25 the use of radioisotopes, affinity labels (such as biotin, avidin, and the like), enzymatic labels (such as horse radish peroxidase, alkaline phosphatase, and the like) fluorescent labels (such as FITC or rhodamine, and the like), paramagnetic atoms, and the like. Procedures
30 for accomplishing such labeling are well-known in the

art, for example, see (Stemberger et al., J. Histochem. Cytochem. 18:315 (1970); Bayer et al., Meth. Enzym. 62:308 (1979); Engval et al., Immunot. 109:129 (1972); Goding, J., Immunol. Meth. 13:215 (1976)). The labeled antibodies of the present invention can be used for in vitro, in vivo, and in situ assays to identify cells or tissues which express a specific peptide.

The above-described antibodies may also be immobilized on a solid support. Examples of such solid supports include plastics such as polycarbonate, complex carbohydrates such as agarose and sepharose, acrylic resins and such as polyacrylamide and latex beads. Techniques for coupling antibodies to such solid supports are well known in the art (Weir et al., "Handbook of Experimental Immunology" 4th Ed., Blackwell Scientific Publications, Oxford, England, Chapter 10 (1986); Jacoby et al., Meth. Enzym. 34, Academic Press, N.Y. (1974)). The immobilized antibodies of the present invention can be used for in vitro, in vivo, and in situ assays as well as in immunochromatography.

Furthermore, one skilled in the art can readily adapt currently available procedures, as well as the techniques, methods and kits disclosed above with regard to antibodies, to generate peptides capable of binding to a specific peptide sequence in order to generate rationally designed antipeptide peptides, for example see Hurby et al., "Application of Synthetic Peptides: Antisense Peptides", In Synthetic Peptides, A User's Guide, W.H. Freeman, NY, pp. 289-307 (1992), and Kaspczak et al., Biochemistry 28:9230-8 (1989).

Anti-peptide peptides can be generated by replacing the basic amino acid residues found in the AUR-1 and/or AUR-2 peptide sequence with acidic residues, while maintaining hydrophobic and uncharged polar groups. For example, lysine, arginine, and/or histidine residues are replaced with aspartic acid or glutamic acid and glutamic acid residues are replaced by lysine, arginine or histidine.

VII. An Antibody Based Method And Kit For Detecting AUR-1 and/or AUR-2.

The present invention encompasses a method of detecting an AUR-1 and/or AUR-2 polypeptide in a sample, comprising: (a) contacting the sample with an above-described antibody, under conditions such that immunocomplexes form, and (b) detecting the presence of said antibody bound to the polypeptide. In detail, the methods comprise incubating a test sample with one or more of the antibodies of the present invention and assaying whether the antibody binds to the test sample. Altered levels of AUR-1 and/or AUR-2 in a sample as compared to normal levels may indicate disease.

Conditions for incubating an antibody with a test sample vary. Incubation conditions depend on the format employed in the assay, the detection methods employed, and the type and nature of the antibody used in the assay. One skilled in the art will recognize that any one of the commonly available immunological assay formats (such as radioimmunoassays, enzyme-linked immunosorbent assays, diffusion based Ouchterlony, or rocket immunofluorescent assays) can readily be adapted

to employ the antibodies of the present invention. Examples of such assays can be found in Chard, "An Introduction to Radioimmunoassay and Related Techniques" Elsevier Science Publishers, Amsterdam, The Netherlands (1986); Bullock et al., "Techniques in Immunocytochemistry," Academic Press, Orlando, FL Vol. 1(1982), Vol. 2 (1983), Vol. 3 (1985); Tijssen, "Practice and Theory of Enzyme Immunoassays: Laboratory Techniques in Biochemistry and Molecular Biology," Elsevier Science Publishers, Amsterdam, The Netherlands (1985).

The immunological assay test samples of the present invention include cells, protein or membrane extracts of cells, or biological fluids such as blood, serum, plasma, or urine. The test sample used in the above-described method will vary based on the assay format, nature of the detection method and the tissues, cells or extracts used as the sample to be assayed. Methods for preparing protein extracts or membrane extracts of cells are well known in the art and can be readily be adapted in order to obtain a sample which is capable with the system utilized.

A kit contains all the necessary reagents to carry out the previously described methods of detection. The kit may comprise: (i) a first container means containing an above-described antibody, and (ii) second container means containing a conjugate comprising a binding partner of the antibody and a label. In another preferred embodiment, the kit further comprises one or more other containers comprising one or more of the

following: wash reagents and reagents capable of detecting the presence of bound antibodies.

Examples of detection reagents include, but are not limited to, labeled secondary antibodies, or in the alternative, if the primary antibody is labeled, the chromophoric, enzymatic, or antibody binding reagents which are capable of reacting with the labeled antibody. The compartmentalized kit may be as described above for nucleic acid probe kits. One skilled in the art will readily recognize that the antibodies described in the present invention can readily be incorporated into one of the established kit formats which are well known in the art.

VIII. Isolation of Compounds Which Interact With AUR-1 and/or AUR-2.

The present invention also relates to a method of detecting a compound capable of binding to a AUR-1 and/or AUR-2 polypeptide comprising incubating the compound with AUR-1 and/or AUR-2 and detecting the presence of the compound bound to AUR-1 and/or AUR-2. The compound may be present within a complex mixture, for example, serum, body fluid, or cell extracts.

The present invention also relates to a method of detecting an agonist or antagonist of AUR-1 and/or AUR-2 activity or AUR-1 and/or AUR-2 binding partner activity comprising incubating cells that produce AUR-1 and/or AUR-2 in the presence of a compound and detecting changes in the level of AUR-1 and/or AUR-2 activity or AUR-1 and/or AUR-2 binding partner activity. The compounds thus identified would produce a change in

activity indicative of the presence of the compound. The compound may be present within a complex mixture, for example, serum, body fluid, or cell extracts. Once the compound is identified it can be isolated using techniques well known in the art.

5 The present invention also encompasses a method of agonizing (stimulating) or antagonizing AUR-1 and/or AUR-2 associated activity in a mammal comprising administering to said mammal an agonist or antagonist to AUR-1 and/or AUR-2 in an amount sufficient to effect
10 said agonism or antagonism. A method of treating diseases in a mammal with an agonist or antagonist of AUR-1 and/or AUR-2 related activity comprising administering the agonist or antagonist to a mammal in an amount sufficient to agonize or antagonize AUR-1
15 and/or AUR-2 associated functions is also encompassed in the present application.

IX. Transgenic Animals.

 A variety of methods are available for the
20 production of transgenic animals associated with this invention. DNA can be injected into the pronucleus of a fertilized egg before fusion of the male and female pronuclei, or injected into the nucleus of an embryonic cell (e.g., the nucleus of a two-cell embryo) following
25 the initiation of cell division (Brinster et al., Proc. Nat. Acad. Sci. USA 82: 4438-4442 (1985)). Embryos can be infected with viruses, especially retroviruses, modified to carry inorganic-ion receptor nucleotide sequences of the invention.

Pluripotent stem cells derived from the inner cell mass of the embryo and stabilized in culture can be manipulated in culture to incorporate nucleotide sequences of the invention. A transgenic animal can be produced from such cells through implantation into a blastocyst that is implanted into a foster mother and allowed to come to term. Animals suitable for transgenic experiments can be obtained from standard commercial sources such as Charles River (Wilmington, MA), Taconic (Germantown, NY), Harlan Sprague Dawley (Indianapolis, IN), etc.

The procedures for manipulation of the rodent embryo and for microinjection of DNA into the pronucleus of the zygote are well known to those of ordinary skill in the art (Hogan *et al.*, *supra*). Microinjection procedures for fish, amphibian eggs and birds are detailed in Houdebine and Chourrout, *Experientia* 47: 897-905 (1991). Other procedures for introduction of DNA into tissues of animals are described in U.S. Patent No., 4,945,050 (Sandford *et al.*, July 30, 1990).

By way of example only, to prepare a transgenic mouse, female mice are induced to superovulate. Females are placed with males, and the mated females are sacrificed by CO₂ asphyxiation or cervical dislocation and embryos are recovered from excised oviducts. Surrounding cumulus cells are removed. Pronuclear embryos are then washed and stored until the time of injection. Randomly cycling adult female mice are paired with vasectomized males. Recipient females are mated at the same time as donor females. Embryos then are transferred surgically. The

procedure for generating transgenic rats is similar to that of mice. See Hammer et al., Cell 63:1099-1112 (1990).

Methods for the culturing of embryonic stem (ES) cells and the subsequent production of transgenic animals by the introduction of DNA into ES cells using methods such as electroporation, calcium phosphate/DNA precipitation and direct injection also are well known to those of ordinary skill in the art. See, for example, Teratocarcinomas and Embryonic Stem Cells, A Practical Approach, E.J. Robertson, ed., IRL Press (1987).

In cases involving random gene integration, a clone containing the sequence(s) of the invention is co-transfected with a gene encoding resistance. Alternatively, the gene encoding neomycin resistance is physically linked to the sequence(s) of the invention. Transfection and isolation of desired clones are carried out by any one of several methods well known to those of ordinary skill in the art (E.J. Robertson, supra).

DNA molecules introduced into ES cells can also be integrated into the chromosome through the process of homologous recombination. Capecchi, Science 244: 1288-1292 (1989). Methods for positive selection of the recombination event (i.e., neo resistance) and dual positive-negative selection (i.e., neo resistance and gancyclovir resistance) and the subsequent identification of the desired clones by PCR have been described by Capecchi, supra and Joyner et al., Nature 338: 153-156 (1989), the teachings of which are incorporated herein. The final phase of the procedure

is to inject targeted ES cells into blastocysts and to transfer the blastocysts into pseudopregnant females. The resulting chimeric animals are bred and the offspring are analyzed by Southern blotting to identify individuals that carry the transgene. Procedures for the production of non-rodent mammals and other animals have been discussed by others. See Houdebine and Chourrout, supra; Pursel et al., Science 244:1281-1288 (1989); and Simms et al., Bio/Technology 6:179-183 (1988).

Thus, the invention provides transgenic, nonhuman mammals containing a transgene encoding a AUR-1 and/or AUR-2 polypeptide or a gene effecting the expression of a AUR-1 and/or AUR-2 polypeptide. Such transgenic nonhuman mammals are particularly useful as an *in vivo* test system for studying the effects of introducing a AUR-1 and/or AUR-2 polypeptide, regulating the expression of a AUR-1 and/or AUR-2 polypeptide (*i.e.*, through the introduction of additional genes, antisense nucleic acids, or ribozymes).

A "transgenic animal" is an animal having cells that contain DNA which has been artificially inserted into a cell, which DNA becomes part of the genome of the animal which develops from that cell. Preferred transgenic animals are primates, mice, rats, cows, pigs, horses, goats, sheep, dogs and cats. The transgenic DNA may encode for a human AUR-1 and/or AUR-2 polypeptide. Native expression in an animal may be reduced by providing an amount of anti-sense RNA or DNA effective to reduce expression of the receptor.

X. Gene Therapy

AUR-1 and/or AUR-2 or its genetic sequences will also be useful in gene therapy (reviewed in Miller, *Nature* 357:455-460, (1992). Miller states that advances have resulted in practical approaches to human gene therapy that have demonstrated positive initial results. The basic science of gene therapy is described in Mulligan, *Science* 260:926-931, (1993).

In one preferred embodiment, an expression vector containing the AUR-1 and/or AUR-2 coding sequence is inserted into cells, the cells are grown *in vitro* and then infused in large numbers into patients. In another preferred embodiment, a DNA segment containing a promoter of choice (for example a strong promoter) is transferred into cells containing an endogenous AUR-1 and/or AUR-2 in such a manner that the promoter segment enhances expression of the endogenous AUR-1 and/or AUR-2 gene (for example, the promoter segment is transferred to the cell such that it becomes directly linked to the endogenous AUR-1 and/or AUR-2 gene).

The gene therapy may involve the use of an adenovirus containing AUR-1 and/or AUR-2 cDNA targeted to a tumor, systemic AUR-1 and/or AUR-2 increase by implantation of engineered cells, injection with AUR-1 and/or AUR-2 virus, or injection of naked AUR-1 and/or AUR-2 DNA into appropriate tissues.

Target cell populations may be modified by introducing altered forms of one or more components of the protein complexes in order to modulate the activity of such complexes. For example, by reducing or inhibiting a complex component activity within target cells, an

abnormal signal transduction event(s) leading to a condition may be decreased, inhibited, or reversed. Deletion or missense mutants of a component, that retain the ability to interact with other components of the protein complexes but cannot function in signal transduction may be used to inhibit an abnormal, deleterious signal transduction event.

Expression vectors derived from viruses such as retroviruses, vaccinia virus, adenovirus, adeno-associated virus, herpes viruses, several RNA viruses, or bovine papilloma virus, may be used for delivery of nucleotide sequences (e.g., cDNA) encoding recombinant AUR-1 and/or AUR-2 protein into the targeted cell population (e.g., tumor cells). Methods which are well known to those skilled in the art can be used to construct recombinant viral vectors containing coding sequences. See, for example, the techniques described in Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y. (1989), and in Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley Interscience, N.Y. (1989). Alternatively, recombinant nucleic acid molecules encoding protein sequences can be used as naked DNA or in reconstituted system e.g., liposomes or other lipid systems for delivery to target cells (See e.g., Felgner et al., *Nature* 337:387-8, 1989). Several other methods for the direct transfer of plasmid DNA into cells exist for use in human gene therapy and involve targeting the DNA to receptors on cells by complexing the plasmid DNA to proteins. See, Miller, *supra*.

In its simplest form, gene transfer can be performed by simply injecting minute amounts of DNA into the nucleus of a cell, through a process of microinjection. Capecchi MR, Cell 22:479-88 (1980). Once recombinant genes are introduced into a cell, they
5 can be recognized by the cells normal mechanisms for transcription and translation, and a gene product will be expressed. Other methods have also been attempted for introducing DNA into larger numbers of cells. These methods include: transfection, wherein DNA is
10 precipitated with CaPO_4 and taken into cells by pinocytosis (Chen C. and Okayama H, Mol. Cell Biol. 7:2745-52 (1987)); electroporation, wherein cells are exposed to large voltage pulses to introduce holes into the membrane (Chu G. et al., Nucleic Acids Res.,
15 15:1311-26 (1987)); lipofection/liposome fusion, wherein DNA is packaged into lipophilic vesicles which fuse with a target cell (Felgner PL., et al., Proc. Natl. Acad. Sci. USA. 84:7413-7 (1987)); and particle bombardment using DNA bound to small projectiles (Yang NS. et al.,
20 Proc. Natl. Acad. Sci. 87:9568-72 (1990)). Another method for introducing DNA into cells is to couple the DNA to chemically modified proteins.

It has also been shown that adenovirus proteins are capable of destabilizing endosomes and
25 enhancing the uptake of DNA into cells. The admixture of adenovirus to solutions containing DNA complexes, or the binding of DNA to polylysine covalently attached to adenovirus using protein crosslinking agents substantially improves the uptake and expression of the

recombinant gene. Curriel DT et al., Am. J. Respir. Cell. Mol. Biol., 6:247-52 (1992).

As used herein "gene transfer" means the process of introducing a foreign nucleic acid molecule into a cell. Gene transfer is commonly performed to enable the expression of a particular product encoded by the gene. The product may include a protein, polypeptide, anti-sense DNA or RNA, or enzymatically active RNA. Gene transfer can be performed in cultured cells or by direct administration into animals. Generally gene transfer involves the process of nucleic acid contact with a target cell by non-specific or receptor mediated interactions, uptake of nucleic acid into the cell through the membrane or by endocytosis, and release of nucleic acid into the cytoplasm from the plasma membrane or endosome. Expression may require, in addition, movement of the nucleic acid into the nucleus of the cell and binding to appropriate nuclear factors for transcription.

As used herein "gene therapy" is a form of gene transfer and is included within the definition of gene transfer as used herein and specifically refers to gene transfer to express a therapeutic product from a cell *in vivo* or *in vitro*. Gene transfer can be performed *ex vivo* on cells which are then transplanted into a patient, or can be performed by direct administration of the nucleic acid or nucleic acid-protein complex into the patient.

In another preferred embodiment, a vector having nucleic acid sequences encoding a AUR-1 and/or AUR-2 is provided in which the nucleic acid sequence is

expressed only in specific tissue. Methods of achieving tissue-specific gene expression as set forth in International Publication No. WO 93/09236, filed November 3, 1992 and published May 13, 1993.

5 In all of the preceding vectors set forth above, a further aspect of the invention is that the nucleic acid sequence contained in the vector may include additions, deletions or modifications to some or all of the sequence of the nucleic acid, as defined above.

10 In another preferred embodiment, a method of gene replacement is set forth. "Gene replacement" as used herein means supplying a nucleic acid sequence which is capable of being expressed *in vivo* in an animal and thereby providing or augmenting the function of an
15 endogenous gene which is missing or defective in the animal.

EXAMPLES

The examples below are non-limiting and are
20 merely representative of various aspects and features of the present invention. The examples below demonstrate the isolation, and characterization of the novel proteins AUR-1 and AUR-2.

Protein kinases are one of the largest
25 families of eukaryotic proteins with several hundred known members. These proteins share a 250-300 amino acid domain that can be subdivided into 12 distinct subdomains that comprise the common catalytic core structure. These conserved protein motifs have recently
30 been exploited using PCR-based cloning strategies

leading to a significant expansion of the known kinases. Multiple alignment of the sequences in the catalytic domain of protein kinases and subsequent phylogenetic analysis permits their segregation into a phylogenetic tree. In this manner, related kinases are clustered into distinct branches or subfamilies including: tyrosine kinases, cyclic-nucleotide-dependent kinases, calcium/calmodulin kinases, cyclin-dependent kinases and MAP-kinases, as well as several other less defined subfamilies.

Initially we set out to identify homologues of CCK4, a receptor that represents a distinct family of tyrosine kinases. Multiple alignments suggested CCK4 was most closely related to ROS and the TRK-family of receptor tyrosine kinases. We designed degenerate primers to conserved sequences within kinase subdomains I and IX of these receptors. Subdomain I is at the N-terminus of the kinase domain and contains the consensus motif GXGXXGXV which is involved in anchoring ATP to the catalytic unit of all classes of kinases. Subdomain IX contains a nearly invariant Asp which acts to stabilize the catalytic loop by bonding to residues in subdomain VIB. This invariant Asp and flanking amino acids are often used in PCR-cloning strategies as it distinguishes tyrosine-kinases (DVWSY/FGI/V) from serine/threonine kinases (DXWA/SXGI/V). Based on comparison of all known protein kinases, we designed degenerate oligonucleotide primers to subdomains I and IX that would pick up only CCK4 and its chicken homologue KLG by PCR.

We designed degenerate primers A and DVW based on conserved residues within the kinase domain of CCK4, to use for identification of novel kinases using polymerase chain reaction (PCR). When applied to HEPM cell ssDNA as a template, multiple copies of CCK4 were isolated as well as a novel DNA fragment (43-43) of 567 bp with homology to other kinases. The novel sequence was most similar to *Drosophila aurora* kinase (GeneBank Accession #X83465) and the clone was designated human Auroral. Using this fragment as a probe, we screened RNAs from a number of colon cancer cell lines and a human multiple tissue Northern blot, demonstrating an apparent selectivity in expression of Auroral in tumor cells.

The Auroral probe was also used to screen a cDNA library constructed from human pancreatic cancer cell line mRNA to isolate overlapping clones spanning the complete open reading frame of Auroral. Of multiple clones isolated, seven corresponded to human Auroral. Two additional faintly hybridizing clones were also isolated during this screen and sequence analysis revealed they corresponded to a related, yet distinct kinase, which we designated human Aurora2.

Recombinant AURORA1 and AURORA2 expressed in COS cells migrated with apparent Mr of 39,000 and 46,000, consistent with their predicted molecular weights of 39264 and 46730 based on their primary amino acid sequence. This analysis confirms the recombinant protein can be stably produced in mammalian cells. Phosphorylation assays to determine target specificity of this putative kinases are ongoing.

Specific immunoreagents were generated in rabbits against peptide sequence from the N-terminal domains of AUR1 and AUR2 have these reagents to localize expression of endogenous and recombinant Auroras within the cells. Furthermore, these reagents can be used in an effort to identify substrates for the AURORAs in order to better understand their normal biological role.

Dominant negative and constitutively active forms of AUR1 and AUR2 will be useful for delineating biologic consequences of either ablation or overexpression of these putative serine/threonine kinases. Initial studies with altered DNA constructs demonstrates that in just 2 hours following infection of NIH3T3 or BALB/3T3 cells with AUR1 or AUR2 retroviral stocks, cells become multinucleated. This phenotype persisted such that 2 days after infection some cells were found to have as many as 20 nuclei. The multinucleated cells typically had increased cytoplasm and diffuse cell boundaries. Immunostaining with both actin and DAPI, confirmed these nuclei were all contained within a single cell, and that the actin cytoskeleton was apparently normal. Ongoing experiments will address the chromosome content and intactness within the nuclei, the number and location of centrosomes, and the general organization of the microtubule network.

Characterization of the long term consequences of expression of AUR1 or AUR2 normal, constitutively active and dominant negative are underway, and particularly whether they induce or reverse cellular

transformation. Stable clones are being isolated that express these recombinant proteins. They will be characterized for growth rate, DNA synthesis, cell-contact inhibition (foci formation), anchorage-independent growth (soft agar assays), and
5 tumorigenicity in nude mice.

What role do the Auroras play in mammalian centrosome replication and segregation? Disruption or deregulation of such functions is known to result in chromosome missegregation, monopolar spindles, and
10 asymeric nuclear division in yeast, drosophila, and amphibians. The homology between human Auroras and the yeast IPL1 and drosophila aurora is striking. We therefore desire to assess if the human auroras are functional equivalents of the yeast IPL1. The yeast IPL1
15 gene is required for high-fidelity chromosome segregation in *Saccharomyces cerevisiae* (Francisco, L., et al., Mol. Cell. Biol. 14:4731-4740, 1994) and a temperature sensitive mutant has been isolated. We plan to determine if various full length and chimeric versions
20 of the human Auroras can complement this temperature sensitive mutant in yeast. The chimeric constructs containing the N-terminal domain of IPL1 and the kinase domain of the human auroras will permit us to determine if the kinase is functionally equivalent, while avoiding
25 potential regulatory or intracellular localization roles mediated by the less well conserved N-terminal domains. Should the human genes compensate for the IPL1 loss of function, we could then use such a line to screen for small molecule inhibitors of the human aurora kinase.

EXAMPLE 1: MOLECULAR CLONING

Total RNAs were isolated using the Guanidine Salts/Phenol extraction protocol of Chomczynski and Sacchi (P. Chomczynski and N. Sacchi, Anal. Biochem. 162, 156 (1987) from normal human prostate, duodenum, 5 ovary, liver, pituitary, brain, thymus, and salivary gland, from human HEPM cells (palatal mesenchyme), from primary human Wilm's tumor and ovarian carcinoma, and from human tumor cell lines originating from colon/rectum (HT29, SW480, SW1463, SW1417, SW837, 10 SW948, SW620, SW403, SW1116, T84, HTC15, LS123, and Caco-2), kidney (CaKi-1, CaKi-2), liver (SK-HEP-1), pancreas (HS766T, ASPC, Capan-1), and breast (MCF7).

These RNAs were used as templates to generate single-stranded cDNAs using the Superscript 15 Preamplification System for First Strand Synthesis kit purchased from GibcoBRL (Life Technologies, U.S.A.; Gerard, GF et al. (1989), FOCUS 11, 66) under conditions recommended by manufacturer. A typical reaction used 10 ug total RNA or 2 ug poly(A)⁺ RNA with 1.5 ug 20 oligo(dT)₁₂₋₁₈ in a reaction volume of 60 ul. The product was treated with RNaseH and diluted to 100 ul with H₂O. For subsequent PCR amplification, 1-4 ul of these sscDNAs were used in each reaction.

Oligonucleotides were synthesized on an 25 Applied Biosystems 394 DNA synthesizer using established phosphoramidite chemistry and were used unpurified after precipitation with ethanol. The degenerate oligonucleotide primers are:

A = 5'-GARTTYGGNGARGTNTTTYTNGC-3' (SEQ ID NO:16)
30 (sense) and

DVW = 5'-AGNACNCCRAANGCCCACACRTC-3' (SEQ ID NO:17)
(antisense).

These primers were derived from the peptide sequences **EFGEVFLA** (SEQ ID NO:18) (sense strand from kinase subdomain I) and **DVW(A/S)FGVL** (antisense strand from kinase subdomain IX), respectively. Degenerate
5 nucleotide residue designations are: N = A, C, G, or T; R = A or G; and Y = C or T. Using CCK4 as a template, these primers produce a product of 567 bp.

A PCR reaction was performed using Primers A
10 and DVW applied to the single-stranded sources listed above. The primers were added at a final concentration of 5 uM each to a mixture containing 10 mM Tris·HCl (pH8.3), 50 mM KCl, 1.5 mM MgCl₂, 200 uM each deoxynucleoside triphosphate, 0.001% gelatin, and 1.5 U
15 AmpliTaq DNA Polymerase (Perkin-Elmer/Cetus), and 1-4 ul cDNA. Following 3 min denaturation at 95°C, the cycling conditions were 94°C for 30 s, 37°C for 1 min, a 2 min ramp to 72°C, and 72°C for 1min for the first 3 cycles, followed by 94°C for 30 s, 50°C for 1 min, and 72°C for
20 1min 45 s for 35 cycles. PCR fragments migrating at between 500-600bp were isolated from 2% agarose gels using GeneClean (Bio101), and T-A cloned into the pCRII vector (Invitrogen Corp. U.S.A.) according to the manufacturer's protocol.

25 Colonies were selected for mini plasmid DNA-preparations using Qiagen columns and the plasmid DNAs were sequenced using cycle sequencing dye-terminator kit with AmpliTaq DNA Polymerase, FS (ABI, Foster City, CA). Sequencing reaction products
30 were run on an ABI Prism 377 DNA Sequencer, and analyzed

using the BLAST alignment algorithm (Altschul, S.F. et al., J. Mol. Biol. 215:403-10). A novel clone (#43-43) was isolated by PCR with primers A and DVW on single-stranded cDNA from human embryonic palatal mesenchyme (HEPM or CRL1486) as a template. This clone
5 was subsequently designated as a fragment of human Auroral.

A lambda ZapII (Stratagene Cloning Systems, La Jolla, CA) cDNA library was constructed using mRNA from a pool of pancreatic carcinoma cell lines as a template
10 for first strand cDNA synthesis. Phage were screened on nitrocellulose filters with the random primed ³²P-labeled insert from p43-43 encoding human Auroral at 2x10⁶ cpm/ml in hybridization buffer containing 6xSSC, 1x Denhardt's reagent, 0.1% SDS, with 0.1mg/ml denatured, fragmented
15 salmon sperm DNA. After overnight hybridization at 65°C, filters were washed in 0.1xSSC, 0.1%SDS at 65°C. Full length cDNA clones were sequenced on both strands using manual sequencing with T7 polymerase and oligonucleotide primers (Tabor and Richardson, 1987, Proc. Natl. Acad.
20 Sci. U.S.A. 84: 4767-71).

EXAMPLE 2: NORTHERN BLOT ANALYSIS

Northern blots containing 2 ug poly A+RNA per lane from 16 different adult human tissues (spleen,
25 thymus, prostate, testis, ovary, small intestine, colonic mucosa, heart, brain, placenta, lung, liver, skeletal muscle, kidney, pancreas, and peripheral blood leukocytes), four different human fetal tissues (brain, lung, liver, and kidney), and 8 human cancer cell lines
30 (HL60, HeLa, K-562, MOLT-4, Raji, SW480, and G361) on a

charge-modified nylon membrane were obtained from Clontech (Palo Alto, CA). Additional Northern blots were prepared by running 10 ug total RNA isolated from human tumor cell lines on a denaturing formaldehyde 1.2% agarose gel and transferring to nylon membranes.

5 Filters were hybridized with random prime [³²P]dCTP-labeled probes synthesized from either the 527 bp insert from human Auroral clone 43-43 or the 1162 bp EcoRI fragment from pSG20, and either the 1kb EcoRI fragment of human Aurora2 clone 11-1A or the 1257 bp
10 BamHI-Not I fragment from pS621. Hybridization was performed at 60°C overnight in 6XSSC, 0.1% SDS, 1X Denhardt's solution, 100 mg/ml denatured herring sperm DNA with 1-2 x 10⁶ cpm/ml of ³²P-labeled DNA probes. The filters were washed in 0.1XSSC/0.1% SDS, 65°C, and
15 exposed overnight on Kodak XAR-2 film.

 A single AUR1 mRNA transcript of approximately 1.4kb was identified, and was found to be most abundant in the thymus and small intestine with weak signals from testis, ovary, colon, placenta, and spleen. Prostate
20 and peripheral blood lymphocytes were negative. Human fetal liver and kidney were also positive, with a weaker signal in fetal lung and no expression in fetal brain (Table)

 A similar analysis of human AUR2 expression
25 showed a more restricted expression profile. A single 2.4 kb AUR2 transcript was detected strongly in the adult testis and thymus, and weakly in [heart, placenta, skeletal muscles] and in fetal liver and kidney whereas the other normal tissue sources were negative (see
30 Table).

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**AURORA 1 and AURORA 2 NORTHERN ANALYSIS IN HUMAN NORMAL
TISSUE AND CANCER CELLS**

	Cell type	Origin	AUR 1	AUR 2
5	Thymus	Normal tissue	5	4
	Fetal liver	Normal tissue	4	2
	Fetal kidney	Normal tissue	4	1
	Lung	Normal tissue	3	0
	Duodenum	Normal tissue	2	1
10	Colon	Normal tissue	2	0
	Fetal lung	Normal tissue	2	0
	Ovary	Normal tissue	2	0
	Testis	Normal tissue	2	2
	Brain	Normal tissue	0	0
15	Cerebellum	Normal tissue	0	0
	Salivary gland	Normal tissue	0	0
	Heart	Normal tissue	0	0
	Liver	Normal tissue	0	0
	Pancreas	Normal tissue	0	0
20	Kidney	Normal tissue	0	0
	Spleen	Normal tissue	0	0
	Stomach	Normal tissue	0	0
	Uterus	Normal tissue	0	0
	Prostate	Normal tissue	0	0
25	Skeletal muscle	Normal tissue	0	0
	Fetal brain	Normal tissue	0	0
	PBLs	Normal tissue	0	0
	Salivary gland	Normal tissue	0	0

	Cell type	Origin	AUR 1	AUR 2
	Placenta	Normal tissue	0	0
	SF-268	CNS tumor	4	ND
	CCRF-CEM	Leukemia	4	ND
	K-562	Leukemia	4	ND
5	HCC-2998	Colon tumor	4	ND
	SW620	Colon tumor	4	2
	KM-12	Leukemia	4	ND
	MCF7/ADR-RES	Breast tumor	4	2
	MDA-N	Breast tumor	4	ND
10	BT-549	Breast tumor	4	ND
	SW480	Colon tumor	4	4
	SW48	Colon tumor	4	ND
	Calu-3	Lung tumor	4	ND
	Calu3	Lung tumor	4	2
15	T47D	Breast tumor	4	2
	A375	Melanoma	4	0
	SF767	CNS tumor	4	0
	SW1417	Colon tumor	4	4
	CaKi2	Kidney tumor	4	0
20	CaKi1	Kidney tumor	4	0
	Caco2	Colon tumor	4	4
	SW1417	Colon tumor	4	0
	T98G	CNS tumor	4	0
	SF-539	CNS tumor	3	ND
25	SK-MEL-2	Melanoma	3	ND
	SK-MEL-5	Melanoma	3	ND

	Cell type	Origin	AUR 1	AUR 2
	R-48	Gastric tumor	3	ND
	RF-1	Gastric tumor	3	ND
	SW948	Colon tumor	3	ND
	AGS	Gastric tumor	3	ND
	HFL1	Normal lung	3	ND
5	OVCAR-8	Ovarian tumor	2	ND
	HT-29	Colon tumor	2	ND
	MDA-MB-231	Breast tumor	2	ND
	MDA-MB-435	Breast tumor	2	ND
	SK-MEL-5	Melanoma	2	ND
10	Kato-3	Gastric tumor	2	ND
	Colo 205	Colon tumor	2	ND
	Colo 320DM	Colon tumor	2	2
	WiDr	Colon tumor	2	ND
	HT-29	Colon tumor	2	ND
15	SNU-C2B	Colon tumor	2	ND
	HTC15	Colon tumor	2	2
	T84	Colon tumor	2	0
	SW948	Colon tumor	2	0
	Daoy	CNS tumor	2	0
20	OVCAR3	Ovary tumor	2	0
	HS766T	Pancreas tumor	2	0
	SW1116	Colon tumor	2	0
	Wilms tumor	Kidney tumor	2	0
25	UO-31	Renal tumor	0	ND

The AUR1 mRNA expression profile in several primary tumors and multiple cell lines of diverse neoplastic origin were determined by Northern analysis and by the semi-quantitative PCR assay using primers from sequences in the AUR1 kinase domain. The results are included in Table. AUR1 transcripts were detected in every tumor line assayed with the highest expression in several human colon cancer cell lines (SW480, Colo320, SW620, SW1417, Caco2, SW12417) and in lung carcinoma (Calu3), breast carcinoma (T47D, MCF7), Melanoma (A375), Kidney carcinoma (CaKi-1, CaKi-2), liver carcinoma (SK-HEP-1), and neural tumors (SF767, T98G). Lesser expression of AUR1 was seen in other colon carcinomas (HTC15, T84, SW948, SW1116, HT29), neural tumors (Daoy), Ovarian carcinoma (Ovcar3, Primary tumor), pancreatic carcinoma (HS766T), and a primary kidney tumor.

The AUR2 expression profile in tumor cell lines was strikingly more restricted than that of AUR1. Strong expression of AUR2 was detected only in colon carcinoma cell lines (Caco2, SW480, SW1417, SW620) whereas weak signals were seen in other colon (HTC15, Colo320), Breast (T47D, MCF7) and lung (Calu3) tumor cell lines. Several other tumor lines had no detectable AUR2 transcripts.

EXAMPLE 3: SEMI-QUANTITATIVE PCR DETECTION OF AURORA1

RNA was isolated from a variety of human cell lines, fresh frozen tissues, and primary tumors. Single stranded cDNA was synthesized from 10 mg of each RNA as described above using the Superscript Preamplification

System (GibcoBRL). These single strand templates were then used in a 35 cycle PCR reaction with two AURORA1-specific oligonucleotides (3476:

5'-TTTGGCTCGGGAGAAGAAAAGCCAT-3' (SEQ ID NO:19), and 3506: 5'-CAATCATCTCTGGGGGCAGGTAGT-3') (SEQ ID NO:20).

5 Reaction products were electrophoresed on 2% agarose gels, stained with ethidium bromide and photographed on a UV light box. The relative intensity of the ~475-bp AURORA1-specific bands were estimated for each sample.

10 EXAMPLE 4: SOUTHERN BLOT ANALYSIS

Genomic DNA was isolated from a variety of transformed human lines (CaCO2, HTC15, LS147T, SKCO4, SW480, SW403, SW620, SW948, SW1417, SW1116, MCF7, BT474) using standard procedures (Maniatis et al.). Cells were
15 trypsinized, washed with PBS and resuspended at ~10⁸ cells/ml in Digestion buffer (100mM NaCl, 10mM Tris pH8, 25 mM EDTA, pH8, 0.5%SDS, 0.1 mg/ml proteinase K). Cells were lysed by incubation at 50°C for 12 hours, followed by extraction with phenol/chloroform and precipitated
20 with an equal volume of 7.5M ammonium acetate and 100% EtOH. DNAs were resuspended in TE buffer.

Approximately 20 micrograms genomic DNA was digested with HindIII or XhoII at 37°C for at least 4 hours before fractionation on 1% agarose gels. The DNA fragments were
25 transferred to nitrocellulose membranes by the capillary transfer method (Southern, EM, J Mol Bio 98:503, 1975) and hybridized with human Aurora1 and Aurora2-specific probes as described for Northern Blot analysis above.

DNAs were restricted with HindIII since both
30 AUR1 and AUR2 cDNAs contain a single site for this

restriction enzyme. AUR1 showed a single 4.3 kb band of equal intensity from all sources suggesting it is a single copy, non-rearranged gene in the multiple tumor types assayed. However, under low stringency conditions, we were able to detect 1.3 kb and 3.2 kb
5 SacI fragments which weakly hybridize to the AUR1 probe. Cloning and sequence analysis reveals this region to encode an intronless AUR1-related pseudogene (termed AUR3), with multiple frame shifts. Furthermore, immediately upstream of the AUR3 pseudogene is a region
10 with complex inverted repeats predicted to form a very stable hairpin loop. AUR3 DNA sequence is homologous to AUR1 beginning from the first nucleotide of the AUR1 cDNA. Immediately upstream of this site, is the predicted hairpin loop of AUR3. We are currently
15 characterizing AUR1 genomic clones to determine if the homology to AUR3 continues upstream from this nucleotide, and whether the AUR1 cDNA includes or be preceded by a similar hairpin loop. AUR2 showed bands at 7.0 kb and 4.3 kb and a faint higher molecular weight
20 band at ~10 kb from all sources. These data suggest AUR2 is also a single copy gene. The multiple bands with seen on blots probed with AUR2 are likely due to the fact that a full length cDNA probe was used.

25

EXAMPLE 5: SEQUENCE ANALYSIS OF cDNA CLONES ENCODING HUMAN AURORA1 and AURORA2

The complete sequence of human Aurora1 and
30 Aurora2 was determined from full length clones of each

isolated from the human pancreatic carcinoma library, from normal human duodenum, and of the partial human Auroral isolated from HEPM cells.

5 The 1,244 bp human Auroral (AUR1_h) nucleotide sequence is shown in SEQ ID NO:1 or SEQ ID NO:2 and contains a single open reading frame encoding a polypeptide of 344 amino acids. The AUR1_h coding region is flanked by a 54 nucleotide 5'-untranslated region and a 132 nucleotide 3'-untranslated region ending with a poly(A) tail.

10 The 2,198 bp human Aurora2 (AUR2_h) nucleotide sequence is shown in SEQ ID NO:1 or SEQ ID NO:2 and contains a single open reading frame encoding a polypeptide of 403 amino acids. The AUR2_h coding region is flanked by a 200 nucleotide 5'-untranslated
15 region and a 768 nucleotide 3'-untranslated region.

The sequences AUR1 and AUR2 cDNAs were sequenced from both a human pancreatic tumor and normal human duodenum, with no sequence differences excepting some likely polymorphic sites. These ambiguities
20 include:

	cDNA	nucleotide	Comment
	AUR1	1174	one clone has poly A inserted
		873	T in all duodenal clones, C in pancreatic tumor
		469	T in one clone, C in all others
		848	G in one clone, A in all others - changes amino acid E to G
5		1097	G in one clone, T in 2 others
		956	G in one clone, A in 4 others
		29	Splice to 103 in 5 clones, no splice (as shown in 5 clones)
	AUR2	349	T in 1 clone, C in multiple others (change amino acid P to L)
10		369	A in 3 clones, G in multiple others (change AA V to I)

The C-terminal portions AUR1 and AUR2 conserve
 all 12 subdomains characteristic of eukaryotic protein
 kinases. This AUR1 and AUR2 kinase domains is preceded
 by a N-terminal domain are 74 and 130 amino acids,
 respectively. Comparison of the AUR1 and AUR2
 nucleotide and deduced amino acid sequences (SEQ ID NO:3
 or SEQ ID NO:4) with the available DNA and protein
 sequence databases indicated that they are unique with
 the exception of several EST sequences sharing high
 sequence identity. They do however have striking
 homology in both the N-terminal and catalytic domains
 with the drosophila aurora and *Saccharomyces cerevisiae*
 IPL1 genes. Furthermore two unpublished database
 entries are likely to be close homologues from *Xenopus*

laevis (p46APK - GB accession #Z17206 and p46BPK - GB accession #Z17207).

The N-terminal domains of Auroras from human, frog, Drosophila and yeast share limited sequence identity. The requiring AUR2 has an abundance of glutamates often present in pairs separated by a single residue.

Comparison of the catalytic domains of these protein reveals AUR1 shares 70% amino acid identity with AUR2, 61% with the Drosophila aurora, and 45% with the yeast IPL1 gene. AUR2 kinase shares 60% amino acid identity with the Drosophila protein and 45% identity to yeast IPL1. Both AUR1 and AUR2 share less than 45% homology with all other known mammalian kinases (the closest being cAMP-dependent protein kinase A) suggesting they are homologues of these drosophila and yeast kinases.

AUR1 and AUR2 both contain a cAMP-dependent protein kinase phosphorylation site (THR232 of AUR1 and THR288 of AUR2) that is conserved in the drosophila and yeast homologues and is a known regulatory site in the cyclin-dependent kinase p34cdc2. AUR2 contains an additional PKA-site at SER342. Both proteins also have multiple Casein kinase II (five and six for AUR1 and AUR2) and protein kinase C (four and ten for AUR1 and AUR2) phosphorylation sites. AUR2 also has a single tyrosine phosphorylation consensus site at TYR334 that is also conserved with the Drosophila aurora, but is not present in AUR1 or yeast IPL1.

Intriguingly, natural mutants of the drosophila aurora AUR_{dm} and yeast IPL1 gene result in

asymmetric nuclear division leading to chromosome missegregation, and atypical, monopolar spindles. This phenotype appears to result from a failure of centrosome separation. The associated microtubule architecture appear unaffected. Natural mutants in both drosophila and yeast target amino acid residues that are strictly conserved between the human Auroras, further supporting they may be functional homologues. The corresponding residues in AUR1 that are found in natural mutants of AUR_dm or IPL1 are GLU125, THR232, PRO312, HIS324. All of these mutations are within the catalytic domain, and notably, one represents the conserved PKA-phosphorylation site. An additional mutation in AUR_dm at ASP47 is at a non-conserved residue in the N-terminal domain.

These findings suggest the catalytic activity may indeed play a central role in the biology of centrosome replication or segregation in lower organisms, and suggest that the human Auroras may play a complementary role in mammalian cells.

EXAMPLE 6: RECOMBINANT EXPRESSION OF AURORA and AURORA2 EXPRESSION VECTOR CONSTRUCTION

Expression constructs were generated by PCR-assisted mutagenesis in which the entire coding domains of Auroral and Aurora2 were tagged on their carboxy-terminal ends with the hemophilus influenza hemagglutinin (HA) epitope YPYDVDPYAS (SEQ ID NO:21) (Pati, 1992). These constructs were introduced into two mammalian expression vectors: pLXSN (Miller, A.D. & Rosman, G.J., Biotechniques 7, 980-988, 1989) for the

generation of virus producing lines; and pRK5 for transient expression analysis. Inserts were designed to be flanked by unique BamHI and NotI sites and were cloned directly into pLXSN or pRK5 at the 5'-BamHI and 3'-NotI sites.

5 The BamHI-NotI full length AUR1 and AUR2 constructs were also ligated into pRS316 (Liu, H. et al, Genetics 132:665-673, 1992). This vector contains a galactose-inducible promoter in a centromeric shuttle vector for expression in *Saccharomyces cerevisiae*.
10 These are to assess if the human genes can complement the related temperature sensitive yeast IPL1 mutant, which is closely related to AUR1. In addition, fusion constructs containing the N-terminal domain of yeast
15 IPL1 fused to the C-terminal kinase domains of AUR1 and AUR2 were generated. These were produced by insertion of an artificial ClaI site at the 5' end of the kinase domains of the kinases, at the conserved Asp-Asp-Phe-Glu sequence.

 Dominant negative AUR1 and AUR2 constructs
20 were also made in both pLXSN and pRK5 by mutation of the invariant Lys (amino acid positions 106 and 162 in AUR1 and AUR2, respectively) to an Met by PCR mutagenesis. The constructs are termed AUR1KM and AUR2KM.
 Constitutively active forms of AUR1 and AUR2 were
25 generated by mutation of the DNA heading the encoding the phosphorylation site (232 and 288) to an Asp resulting in AUR1TD and AUR2TD.

 Expression constructs in both pLXSN and pRK5
30 were also made containing just the N-terminal, non-catalytic domain of AUR1 and AUR2. These were

generated by PCR from the parental constructs and contain the N-terminal 77 amino acids of AUR1 and 132 amino acids of AUR2.

The entire AUR1 and AUR2 open reading frames (no HA-tag) excluding the initiating methionines were generated by PCR and ligated into pGEX vector for
5 bacterial production of GST-fusion proteins for immunization of rabbits for antibody production.

10 **EXAMPLE 7: GENERATION OF VIRUS PRODUCING AURORA CELL LINES**

To generate high-titer virus stocks, pLXSN recombinant constructs containing either AUR1 or AUR2 genes were transfected into an amphotropic helper cell line PA317 using CaCl_2 -mediated transfection. After
15 selection on G418, the cells were plated on normal media without G418 (500 ug/ml). Supernatants from resistant cells were used to infect the ecotropic helper cell line GP+E86, and cells again selected on G418. Resistant cells were again taken off G418, and the supernatants
20 harvested every 8-12 hours and pooled as virus stock (Redemann, N., Holzmann, B., Wagner, E.F., Schlessinger, J., & Ullrich, A., Mol. Cell. Biol. 12, 491-498, 1992). Viral stock titers were typically $\sim 10^6/\text{ml}$.

25 **EXAMPLE 8: RETROVIRAL INFECTION OF NIH-3T3 CELLS WITH AURORAS**

NIH-3T3, and BALB/3T3 cells were grown in 100 mm plates with DMEM (Gibco) containing 10% fetal calf serum (FCS). The cells were superinfected with the AUR1
30 and AUR2 retrovirus by adding approximately 3 ml viral

supernatant to 15 ml culture media for approximately 24 hours. Cells expressing the retroviral constructs were then selected by growth in DMEM/10% FCS supplemented with 500 ug/ml G418.

5 **EXAMPLE 9: GENERATION OF AURORA-SPECIFIC IMMUNOREAGENT**

AURORA-specific immunoreagents were raised in rabbits against KLH-conjugated synthetic peptides corresponding to either the N-terminal region of AUR2 (¹⁰⁴SAPENNPEEQLASK¹¹⁷) (SEQ ID NO:22) and
10 (⁹⁰RPLNNTQKSKQPL¹⁰²) (SEQ ID NO:23) or the N-terminus or N-terminal domain of human AUR1 (¹MAQKENSYPWPYG¹³) (SEQ ID NO:24) and (⁵³PGQKVMENSSGTP⁶⁵) (SEQ ID NO:25). Additional immunoreagents were generated by immunizing rabbits with the bacterially expressed full length AUR1
15 and AUR2 GST-fusion proteins.

EXAMPLE 10: TRANSIENT EXPRESSION OF AURORAS IN MAMMALIAN CELLS

The pRK5 expression plasmids (10ug DNA/100 mm
20 plate) containing the HA-tagged AUR1 and AUR2 genes were introduced into COS and 293 cells with lipofectamine (Gibco BRL). After 72 hours, the cells were harvested in 0.5 ml solubilization buffer (20 mM HEPES pH7.35, 150 mM NaCl, 10% glycerol, 1% Triton X-100, 1.5 mM MgCl₂, 1
25 mM EGTA, 2 mM phenylmethylsulfonyl fluoride, 1 µg/ml aprotinin). Sample aliquots were resolved by SDS polyacrylamide gel electrophoresis (PAGE) on 15%acrylamide/0.5% bis-acrylamide gels and electrophoretically transferred to nitrocellulose.
30 Non-specific binding was blocked by preincubating blots

in Blotto (phosphate buffered saline containing 5% w/v non-fat dried milk and 0.2% v/v nonidet P-40 (Sigma)), and recombinant protein was detected using a murine Mab to the HA decapeptide tag. Alternatively, recombinant protein can be detected using various AUR1- or
5 AUR2-specific antisera.

**EXAMPLE 11: MYELIN BASIC PROTEIN IS AN ARTIFICIAL
SUBSTRATE FOR AURORA1 AND AURORA2 KINASE**

Method

10 Human colorectal adenocarcinoma SW480 cells were cultured in RPMI 1640 plus 10% fetal bovine serum, L-glutamine, penicillin and streptomycin. Confluent cultures of SW480 cells were washed three times with ice cold phosphate buffered saline (PBS) and then were
15 scraped into 1 ml of ice cold PBS. The cells were centrifuged at 1,000 rpm at 4°C, the PBS aspirated away, and the resulting cell pellet stored at -80°C. The pellets from three 15 cm plates were thawed on ice and resuspended in a total of 1 ml of kinase lysis buffer 50
20 mM HEPES pH 7.4, 100 mM KCl, 25 mM NaF, 1 mM NaVO₃, 0.5% NP40, 1 mM DTT, 2 ug/ml aprotinin, and 1 ug/ml leupeptin) and were rotated gently for 20 minutes at 4°C. The samples were then centrifuged at 10,000 x g for 10 minutes at 4°C and the resulting supernatant was
25 transferred to a clean 1.5 ml centrifuge tube and stored or kept on ice. The protein concentration was determined by Bradford analysis. One milligram of total protein was pre-cleared with 10 µl of protein A-Sepharose (Boehringer) for 15 minutes at 4°C followed by
30 the addition of 2 µl of either rabbit pre-immune serum,

affinity purified auroral peptide antisera, affinity purified auroral peptide antisera plus 6 μ g of competing auroral peptide, affinity purified aurora2 peptide antisera, or, affinity purified aurora2 peptide antisera plus 6 μ g of competing aurora2 peptide and incubated for 5 30 minutes at 4°C. Subsequently, 10 μ l of protein A-sepharose was added and the incubation was continued for an additional 45 minutes at 4°C. The tubes were briefly centrifuged to pellet the antibody-protein A-sepharose complex and the resulting supernatant was aspirated off. 10 The antibody-protein A-sepharose pellet was washed twice with 0.5 ml of kinase lysis buffer followed by a wash with 0.5 ml of kinase buffer (20 mM HEPES pH 7.4, 125 mM KCl, 10 mM MgCl₂, 1 mM NaF, 1 mM NaVO₃, and 1 mM DTT). The antibody-protein A-sepharose pellet was resuspended 15 in 20 μ l of kinase buffer containing 5 uCi of [γ -³²P] ATP and 0.5 mg/ml myelin basic protein (Sigma), incubated for 20 minutes at 37°C after which 10 μ l of protein sample buffer 200 mM Tris-Cl pH 6.8, 40% glycerol, 730 mM B-mercaptoethanol, 0.4% SDS, and 0.05% Bromophenol 20 Blue) was added. The tubes were mixed well and incubated for 5 minutes at 100°C. The samples were resolved on an 18% SDS polyacrylamide gel and visualized by autoradiography.

Results

25 Auroral and aurora2 immunocomplexes were able to phosphorylate myelin basic protein. When competing peptide was used in the immunoprecipitations neither auroral nor aurora2 antisera immunocomplexes were able to phosphorylate myelin basic protein more than the pre- 30 immune sera control. This suggests that the kinase

activity observed is due to auroral and 2 and not to other proteins present in the immunocomplex.

This observation will allow for the purification of active auroral and 2 kinase by using myelin basic protein as a substrate to follow kinase activity. It also will allow the development of an in vitro kinase assay using recombinant auroral and 2 proteins. Furthermore an auroral and 2 in vitro kinase assay will allow one to screen small molecule collections for inhibitors of the auroral and 2 kinases by measuring the inhibition of phosphorylation of myelin basic protein.

The invention illustratively described herein suitably may be practiced in the absence of any element or elements, limitation or limitations which is not specifically disclosed herein. The terms and expressions which have been employed are used as terms of description and not of limitation, and there is no intention that in the use of such terms and expressions of excluding any equivalents of the features shown and described or portions thereof, but it is recognized that various modifications are possible within the scope of the invention claimed. Thus, it should be understood that although the present invention has been specifically disclosed by preferred embodiments and optional features, modification and variation of the concepts herein disclosed may be resorted to by those skilled in the art, and that such modifications and variations are considered to be within the scope of this invention as defined by the appended claims.

Those references not previously incorporated herein by reference, including both patent and non-patent references, are expressly incorporated herein by reference for all purposes. Other embodiments are within the following claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Mossie, Kevin G.
Plowman, Gregory D.
- (ii) TITLE OF INVENTION: Human Aurora Application
- 5 (iii) NUMBER OF SEQUENCES: 6
- (iv) CORRESPONDENCE ADDRESS:
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- 10 (B) STREET: 633 West Fifth Street, Suite 4700
- (C) CITY: Los Angeles
- (D) STATE: CA
- (E) COUNTRY: USA
- (F) ZIP: 90071-2066
- (v) COMPUTER READABLE FORM:
- 15 (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
- 20 (A) APPLICATION NUMBER: US
- (B) FILING DATE:
- (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
- (A) NAME: Warburg, Richard J.
- 25 (B) REGISTRATION NUMBER: 32,327
- (ix) TELECOMMUNICATION INFORMATION:
- (A) TELEPHONE: (213) 489-1600
- (B) TELEFAX: (213) 955-0440
- (C) TELEX: 67-3510
- 30 (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1244 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- 35 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2198 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

5 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

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81

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20 (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 344 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

25 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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				20					25					30			
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			35					40					45				
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10	Pro	Asp	Ile	Leu	Thr	Arg	His	Phe	Thr	Ile	Asp	Asp	Phe	Glu	Ile	Gly	
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30	Ala	Pro	Ser	Leu	Arg	Arg	Lys	Thr	Met	Cys	Gly	Thr	Leu	Asp	Tyr	Leu	
	225					230					235					240	
	Pro	Pro	Glu	Met	Ile	Glu	Gly	Arg	Met	His	Asn	Glu	Lys	Val	Asp	Leu	
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83

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 275 280 285
 5 Asp Leu Lys Phe Pro Ala Ser Val Pro Thr Gly Ala Gln Asp Leu Ile
 290 295 300
 Ser Lys Leu Leu Arg His Asn Pro Ser Glu Arg Leu Pro Leu Ala Gln
 305 310 315 320
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 325 330 335
 10 Pro Ser Ala Leu Gln Ser Val Ala
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(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 403 amino acids
 15 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

20 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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 1 5 10 15
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 35 40 45
 Cys Pro Ser Asn Ser Ser Gln Arg Val Pro Leu Gln Ala Gln Lys Leu
 50 55 60
 30 Val Ser Ser His Lys Pro Val Gln Asn Gln Lys Gln Lys Gln Leu Gln
 65 70 75 80
 Ala Thr Ser Val Pro His Pro Val Ser Arg Pro Leu Asn Asn Thr Gln

84

	85								90				95			
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	Glu	Leu	Ala	Ser	Lys	Gln	Lys	Asn	Glu	Glu	Ser	Lys	Lys	Arg	Gln	Trp
			115					120					125			
5	Ala	Leu	Glu	Asp	Phe	Glu	Ile	Gly	Arg	Pro	Leu	Gly	Lys	Gly	Lys	Phe
	130						135					140				
	Gly	Asn	Val	Tyr	Leu	Ala	Arg	Glu	Lys	Gln	Ser	Lys	Phe	Ile	Leu	Ala
	145					150					155					160
10	Leu	Lys	Val	Leu	Phe	Lys	Ala	Gln	Leu	Glu	Lys	Ala	Gly	Val	Glu	His
					165					170					175	
	Gln	Leu	Arg	Arg	Glu	Val	Glu	Ile	Gln	Ser	His	Leu	Arg	His	Pro	Asn
				180					185					190		
	Ile	Leu	Arg	Leu	Tyr	Gly	Tyr	Phe	His	Asp	Ala	Thr	Arg	Val	Tyr	Leu
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15	Ile	Leu	Glu	Tyr	Ala	Pro	Leu	Gly	Thr	Val	Tyr	Arg	Glu	Leu	Gln	Lys
	210						215					220				
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	225					230					235					240
20	Ala	Asn	Ala	Leu	Ser	Tyr	Cys	His	Ser	Lys	Arg	Val	Ile	His	Arg	Asp
				245						250					255	
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25	Ala	Asp	Phe	Gly	Trp	Ser	Val	His	Ala	Pro	Ser	Ser	Arg	Arg	Thr	Thr
		275						280					285			
	Leu	Cys	Gly	Thr	Leu	Asp	Tyr	Leu	Pro	Pro	Glu	Met	Ile	Glu	Gly	Arg
		290				295						300				
	Met	His	Asp	Glu	Lys	Val	Asp	Leu	Trp	Ser	Leu	Gly	Val	Leu	Cys	Tyr
	305					310					315					320
30	Glu	Phe	Leu	Val	Gly	Lys	Pro	Pro	Phe	Glu	Ala	Asn	Thr	Tyr	Gln	Glu
					325					330					335	
	Thr	Tyr	Lys	Arg	Ile	Ser	Arg	Val	Glu	Phe	Thr	Phe	Pro	Asp	Phe	Val
				340					345					350		
35	Thr	Glu	Gly	Ala	Arg	Asp	Leu	Ile	Ser	Arg	Leu	Leu	Lys	His	Asn	Pro
		355						360					365			

85

Ser Gln Arg Pro Met Leu Arg Glu Val Leu Glu His Pro Trp Ile Thr
370 375 380

Ala Asn Ser Ser Lys Pro Ser Asn Cys Gln Asn Lys Glu Ser Ala Ser
385 390 395 400

Lys Gln Ser

5

What is claimed is:

CLAIMS

1. An isolated, enriched or purified nucleic acid molecule encoding AUR-1 and/or AUR-2 polypeptide.
2. The nucleic acid molecule of claim 1
5 wherein said nucleic acid is human nucleic acid.
3. The nucleic acid molecule of claim 1 wherein said molecule encodes at least 25 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.
- 10 4. The nucleic acid molecule of claim 1 wherein said molecule encodes at least 50 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.
- 15 5. The nucleic acid molecule of claim 1 wherein said molecule encodes at least 100 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.
- 20 6. The nucleic acid molecule of claim 1 wherein said molecule encodes at least 200 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

7. The nucleic acid molecule of claim 1 wherein said molecule encodes at least 300 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

8. A nucleic acid probe for the detection of
5 nucleic acid encoding an AUR-1 and/or AUR-2 polypeptide in a sample.

9. The probe of claim 8 wherein said polypeptide comprises at least 25 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ
10 ID NO:4.

10. The probe of claim 8 wherein said polypeptide comprises at least 50 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

11. The probe of claim 8 wherein said
15 polypeptide comprises at least 100 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

12. The probe of claim 8 wherein said
20 polypeptide comprises at least 200 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

13. The probe of claim 8 wherein said polypeptide comprises at least 300 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

14. A recombinant nucleic acid molecule
5 encoding an AUR-1 and/or AUR-2 polypeptide and a vector or promoter effective to initiate transcription in a host cell.

15 15. A recombinant nucleic acid molecule comprising a transcriptional region functional in a cell, a sequence complementary to an RNA sequence encoding an AUR-1 or AUR-2 polypeptide and a transcriptional termination region functional in a cell.

16. An isolated, enriched or purified AUR-1 or AUR-2 polypeptide.

15 17. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide comprises at least 25 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

20 18. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide comprises at least 50 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

19. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide comprises at least 100 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

20. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide comprises at least 200 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

21. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide comprises at least 300 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

22. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide is isolated, purified, or enriched from a mammal.

23. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide is isolated, purified, or enriched from a human.

24. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide is an AUR-1 polypeptide.

25. The AUR-1 or AUR-2 polypeptide of claim 16 wherein said polypeptide is an AUR-2 polypeptide.

26. An antibody having specific binding affinity to an AUR-1 or AUR-2 polypeptide.

27. The antibody of claim 26 wherein said AUR-1 or AUR-2 polypeptide comprises at least 3 contiguous amino acids of the amino acid sequence shown
5 in SEQ ID NO:3 or SEQ ID NO:4.

28. The antibody of claim 26 wherein said AUR-1 or AUR-2 polypeptide comprises at least 4 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

10 29. The antibody of claim 26 wherein said AUR-1 or AUR-2 polypeptide comprises at least 25 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

15 30. The antibody of claim 26 wherein said AUR-1 or AUR-2 polypeptide comprises at least 50 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

20 31. The antibody of claim 26 wherein said AUR-1 or AUR-2 polypeptide comprises at least 100 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

32. The antibody of claim 26 wherein said polypeptide is isolated, purified, or enriched from a mammal.

33. The antibody of claim 26 wherein said polypeptide is isolated, purified, or enriched from a
5 human.

34. The antibody of claim 26 wherein said polypeptide is an AUR-1 polypeptide.

35. The antibody of claim 26 wherein said polypeptide is an AUR-2 polypeptide.

10 36. A hybridoma which produces an antibody having specific binding affinity to a AUR-1 and/or AUR-2 polypeptide.

37. The hybridoma of claim 36 wherein said AUR-1 or AUR-2 polypeptide comprises at least 25
15 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

38. The hybridoma of claim 36 wherein said AUR-1 or AUR-2 polypeptide comprises at least 50
contiguous amino acids of the amino acid sequence shown
20 in SEQ ID NO:3 or SEQ ID NO:4.

39. The hybridoma of claim 36 wherein said AUR-1 or AUR-2 polypeptide comprises at least 100

contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

40. The hybridoma of claim 36 wherein said AUR-1 or AUR-2 polypeptide comprises at least 200 contiguous amino acids of the amino acid sequence shown
5 in SEQ ID NO:3 or SEQ ID NO:4.

41. The hybridoma of claim 36 wherein said AUR-1 or AUR-2 polypeptide comprises at least 300 contiguous amino acids of the amino acid sequence shown in SEQ ID NO:3 or SEQ ID NO:4.

10 42. The hybridoma of claim 36 wherein said polypeptide is isolated, purified, or enriched from a mammal.

43. The hybridoma of claim 36 wherein said polypeptide is isolated, purified, or enriched from a
15 human.

44. The hybridoma of claim 36 wherein said polypeptide is an AUR-1 polypeptide.

45. The hybridoma of claim 44 wherein said polypeptide is an AUR-2 polypeptide.

INTERNATIONAL SEARCH REPORT

International Application No

PC./US 96/18859

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/54 C12N15/85 C12N9/12 C12N5/12 C07K16/40
C12Q1/68 G01N33/577 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K C12Q G01N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EMBL SEQUENCE DATABASE, 1 October 1995, HEIDELBERG, BRD, XP002029128 L. HILLIER ET AL.: "yq61f04.s1 Homo sapiens cDNA clone 200287 3'" accession no. R97724 ---	8-11
X	EMBL SEQUENCE DATABASE, 1 October 1995, HEIDELBERG, BRD, XP002029129 L. HILLIER ET AL.: "yq60d11.s1 Homo sapiens cDNA clone 200181 3' similar to SP:S34642" accession no. R97912 --- -/-	8-11

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

9 April 1997

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 96/18859

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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A	<p>WO 94 23039 A (CANCER RES INST ROYAL ;MARSHALL CHRISTOPHER JOHN (GB); ASHWORTH AL) 13 October 1994 cited in the application see the whole document ---</p>	1-45
P,A	<p>WO 96 34985 A (SUGEN INC) 7 November 1996 see the whole document -----</p>	1-45

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 96/18859

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